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Descripti n

The present invention relates to a human granulocyte colony stimulating factor. More particularly, the present invention relates to a human chromosomal gene coding for a polypeptide having the activity of a colony stimulating factor (hereinafter abbreviated as CSF) which is a specific stimulating factor necessary for the principal purpose of forming colonies of human granulocytic cells. The present invention also relates to a recombinant vector inserted said gene, a transformant containing said vector and a process for producing a glycoprotein having the CSF activity.

When bone marrow cells as target cells and kidney cells or fetal cells were cultured by the double-layer soft agar cultivation method, with the bone marrow cells being in the upper layer and the kidney or fetal cells in the lower layer, part of the cells in the upper layer grew and differentiated to form colonies of neutrophilic granulocytes (hereunder simply referred to as granulocytes) or monocytic macrophages. This observation has led to the assumption of the presence in vivo of factors which promote the formation of colonies [Pluznik and Sach, J. Cell. Comp. Physiol., 66, 319 (1965); and Bradley and Metcalf, Aust. J. Exp. Biol. Med. Sci., 44, 287 (1966)].

These factors which are collectively referred to as CSF are known to be produced by cells, such as T-cells, monocytic macrophages, fibroblasts and endothelial cells, which normally are distributed extensively in vivo. Among subclasses of CSF are included: granulocyte-monocytic macrophage CSF (abbreviated as GM-CSF) which act on the stem cells of granulocytes or monocyte macrophages in such a manner that they stimulate the growth of such stem cells and induce their differentiation to form colonies of granulocytes or monocytic macrophages; monocytic macrophage CSF (abbreviated as M-CSF) which is principally capable of forming colonies of monocytic macrophages; multipotent CSF (abbreviated as multi-CSF) which acts on less differentiated multipotent stem cells; and granulocyte CSF (abbreviated as G-CSF) of the typ contemplated by the present invention which is principally capable of forming granulocytic colonies. It has recently been held that the stages of differentiation of target cells differ from one subclass of CSF to another [Asano, Taisha - Metabolism and Disease, 22, 249 (1985); and Yunis et al., "Growth and Maturation Factors", edited by Guroff, John Wiley & Sons, NY, vol. 1, 209 (1983)].

Therefore, purifying the individual CSF subclasses and making a closer study of their chemical and biological properties are very important for the purpose of estimating the hematopoietic mechanisms and analyzing the pathomorphological aspects of various hematological diseases. The biological actions of G-CSF that are drawing increasing attention of researchers are their capabilities of inducing the differentiation of bone marrow leukemic cells and enhancing the functions of mature granulocytes, and much promise has been held in the potential clinical utility of G-CSF in the fields of treating and preventing leukemia.

The attempts heretofore made to isolate and purify G-CSF are based on the method of cell cultivation wherein G-CSF is isolated from the supernatant of a cell culture, but homogeneous G-CSF has yet to b produced in large quantities by this method because G-CSF can only be produced in low concentration and complex purification procedures are required to obtain a trace amount of G-CSF from a large volume of culture solution. Therefore, it has been strongly desired to achieve mass production of G-CSF by recombinant DNA technology.

One object of the present invention is to provide a human chromosomal gene encoding a polypeptide having human G-CSF activity.

Another object of the present invention is to provide a recombinant vector having inserted said gene.

Still another object of the present invention is to provide a transformant which has been produced by transforming a host with said recombinant vector.

A further object of the present invention is to provide a process for producing a glycoprotein having th human G-CSF activity.

The glycoprotein has a sugar chain portion and a polypeptide which is represented by all or part of th amino acid sequence shown below:

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln
Ser Phe Leu Leu Lys Cys Leu Glu Gln Val Arg
Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu
Lys Leu (Val Ser Glu)$_m$ Cys Ala Thr Tyr Lys Leu
Cys His Pro Glu Glu Leu Val Leu Leu Gly His
Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser
Cys Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys
Leu Ser Gln Leu His Ser Gly Leu Phe Leu Tyr
Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln
Leu Asp Val Ala Asp Phe Ala Thr Thr Ile Trp
Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala
Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe
Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly Val
Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu
Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln
Pro

(where m is 0 or 1).

Fig. 1 shows the sequences of three different probes, IWQ, A and LC;

Fig. 2 shows the nucleotide sequence of a pHCS-1 insert;

Fig. 3 shows the nucleotide sequence of a cDNA insert in pBRG4;

Fig. 4 shows the nucleotide sequence of a cDNA insert in pBRV2;

Fig. 5 shows the nucleotide sequence of a human chromosomal gene coding for human G-CSF;

Fig. 6 shows the restriction enzyme cleavage sites of the human chromosomal gene coding for human G-CSF;

Fig. 7 shows schematically the structure of pMLCE3α;

Fig. 8 shows schematically the structure of pTNCE3α; and

Fig. 9 shows schematically the structures of pD26SVCE3α and pDRCE3α.

The human chromosomal gene of the present invention contains a nucleotide sequence that takes part in transcriptional control and it also contains all or part of the nucleotide sequence shown in Fig. 5.

A chromosomal gene may be obtained by first preparing from human cells a set of recombinants containing a human chromosomal gene (the set is hereunder referred to as a human chromosomal gene library), then subjecting said human chromosomal gene library to screening by known procedures.

The human chromosomal gene may be supplied from any type of human cells such as cells extracted from the liver or kidney or cultured cells such as tumor cells. A human chromosomal gene library may be prepared from human cells by any of the known methods [see Maniatis et al., Cell, 15, 687 (1978); and Maniatis et al., Molecular Cloning, Cold Spring Harbor Laboratory, p. 269 ff. (1982)], which are illustrated below:

extract a human chromosomal DNA from such sources as human fetal liver with phenol or other appropriate chemicals; digest the extracted DNA partially or completely with an appropriate restriction enzyme to obtain a DNA fragment of an appropriate length; insert the DNA fragment into a λ-phage vector DNA fragment with a T4 DNA ligase or other appropriate ligases, with a linker containing the restriction site for an appropriate enzyme such as EcoRI being optionally attached; subsequently, obtain λ-phage particles by in vitro packaging method and transform host cells such as E. coli with the resulting λ-phage particles.

Examples of th λ-phag usable as the vector in the above procedures includ Charon 4A and EMBL-3 and EMBL-4.

Screening for the phage harboring the desired gen from a human chromosomal gene library may b performed by, for example, the following procedures.

A human G-CSF protein obtained from human G-CSF producing cells is purified and a partial amino acid sequence of the protein is determined. In a separate step, messenger RNA (mRNA) is xtracted from the G-CSF producing cells and a complementary DNA to the mRNA (cDNA) is synthesized. A group of recombinant DNAs which contain the thus obtained cDNA (cNDA library) is thereafter prepared. Screening of the cDNA library is conducted using an olygonucleotide probe which has been chemically synthesized on the basis of the aforementioned amino acid sequence (see "Molecular Cloning", ibid.)

A DNA fragment which has all or part of the full-length DNA encoding a human G-CSF polypeptid is obtained from the cDNA clones. The human chromosomal gene library already described in this specification is screened by the plaque hybridization method with this DNA fragment being used as a probe [Benton and Davis, Science, 196, 180 (1977)].

The fragment harboring the thus cloned chromosomal gene coding for the polypeptide having the human G-CSF activity may be re-inserted in an appropriate vector DNA for the purpose of transforming other eukaryotic host cells. By introducing an appropriate promoter and an expression-associated sequence into the vector, the gene can be expressed in an individual host cell.

Illustrative host cells derived from mammalian cells include COS cells, Chinese hamster ovary (CHO) cells, C-127 cells and Hela cells. An illustrative vector that may be used to transform these cells is pdBPV-1 [see Sarver et al.; Proc. Natl. Acad. Sci., USA, 79, 7147 (1982)]. The vectors used to transform these cells contain origin, selection marker, a promoter preceding in position the gene to be expressed, RNA splicing site, polyadenylation signal, etc.

Illustrative promoters that may be used for gene expression in mammalian cells include the promoters of a retrovirus, polyoma virus, adenovirus, simian virus 40 (SV40), etc. If the promoter of SV40 is used, th desired gene expression may be readily achieved in accordance with the method of Mulligan et al. described in Nature, 277, 108 (1979).

Illustrative origins that can be used include those derived from SV40, polyoma virus, adenovirus, bovine papilloma virus (BPV), etc. Illustrative selection markers that can be used include the phosphotransferase APH (3') II or I (neo) gene, thymidine kinase (TK) gene, E. coli xanthineguanine phosphoribosyltransferase (Ecogpt) gene, dihydrofolate reductase (DHFR) gene, etc.

In order to obtain polypeptides having the human G-CSF activity from the above listed host-vector systems, the following procedures may be used: the gene coding for the peptide having the human G-CSF activity is inserted at a suitable site in one of the vectors mentioned above; the host cell is transformed with the resulting recombinant DNA; and the obtained transformants are cultured. The desired polypeptide may be isolated and purified from the cell or culture solution by any one of the known techniques.

Eukaryotic genes are generally held to exhibit polymorphysm as is known for the case of the human interferon gene [see Nishi et al., J. Biochem., 97, 153 (1985)] and this phenomenon may cause substitution of one or more amino acids or a change in the nucleotide sequence but no change in the amino acid sequence at all.

It is also known that each allele of a chromosome may have a different nucleotide sequence [see, for example, Weissmann et al., Phil. Trans. R. Soc. Lond. B299, 7 (1982)].

The G-CSF activity may also be possessed by a polypeptide which is deficient of one or more of the amino acids in the amino acid sequence or which has such amino acids added thereto, or a polypeptide which has one or more of these amino acids replaced by one or more amino acids. It is also known that a polypeptide obtained by converting each of the cysteine codons in the human interleukin-2 (IL-2) gene to a serine codon has the activity of interleukin-2 [Wang et al., Science, 224, 1431 (1984)]. Therefore, so long as the polypeptides, either naturally occurring or chemically synthesized, have the human G-CSF activity, all of the genes that code for these polypeptides, recombinant vectors containing these genes, transformants obtained by such recombinant vectors, and the polypeptides or glycoproteins that are obtained by cultivating such transformants are included within the scope of the present invention.

Hereunder outlined are the processes for producing the chromosomal gene of the present invention coding for a polypeptide having the human G-CSF activity, a recombinant vector having said gene and a transformant having this recombinant vector, and a process for producing a glycoprotein having the human G-CSF activity.

(1) Probe preparation

A homogen ous human CSF prot in was purified from th supernatant of a culture of a tumor cell line, CHU-2 (deposited at Coll ction Nationale de Cultures de Microorganismes, or C.N.C.M., under Accession Number I-483), and its amino acid sequenc from the N terminus was determined. Fragm nts were obtained by decomposition with bromocyan and treatment with trypsin and th amino sequences of these

fragments were also determined [Example 3(i), (ii) and (iii)].

From the determined amino acid sequences, three nucleotide probes, (A), (LC) and (IWQ), having the sequences shown in Fig. 1 were synthesized (Example 4). Probe (A) was of the mixed type composed of 14 successive nucleotides. Probe (IWQ) was composed of 30 successive nucleotides with deoxyinosine and was a probe of the type used in the cloning of the human cholecystokinin gene [Takahashi et al., Proc. Natl. Acad. Sci., USA, 82, 1931 (1985)]. Probe (LC) was a 24-nucleotide probe that was synthesized from the nucleotides at 32 - 39 positions from the N terminus of the amino acid sequence shown in Example 3(i) on the basis of the nucleotide sequence shown in Fig. 3.

Chemical synthesis of nucleotides can be achieved by applying the improved phosphotriester method to the solid phase method and has been reviewed by Narang [Tetrahedron, 39, 3-22 (1983)]. Thes nucleotides can also be synthesized using an automatic DNA synthesizer (Applied Biosystems).

Probes based on amino acid sequences at positions other than those in the above-mentioned probes may also be used.

(2) Construction of cDNA library

CHU-2 cells were homogenized after addition of a guanidine thiocyanate solution and the total RNA was obtained by CsCl density gradient centrifugation.

Poly(A$^+$) RNA was isolated from the total RNA by column chromatography on oligo(dT)-cellulose. Thereafter, a single-stranded cDNA was synthesized with a reverse transcriptase, and RNase H and E. coli DNA polymerase I were added to obtain a double-stranded cDNA. A dC chain was attached to the obtained double-stranded cDNA, which was joined to a vector, pBR322, to which a dG chain had been attached at the Pst I cleavage site. The resulting recombinant DNA was used to transform a strain of E. coli, X1776, and a pBR322-line cDNA library was constructed (Examples 5 and 6).

In a similar manner, the double-stranded cDNA was joined to the λgt10 vector with the EcoRI linker and λ-phage line cDNA library was constructed (Example 7).

(3) Screening

Recombinants derived from the pBR322-line cDNA library were fixed on Whatmann 541 filter paper and a single clone could be selected by colony hybridization with ^{32}P-labelled probe (IWQ). Further study with the Southern blotting method [Southern, J. Mol. Biol., 98, 503 (1975)] showed that this clone also hybridized with probe (A). The nucleotide sequence of this clone was determined by the dideoxy method [Sanger, Science, 214, 1205 (1981)].

The nucleotide sequence of the obtained cDNA insert is shown in Fig. 2, from which one can see that this insert consisted of 308 base pairs including probes (IWQ) and (A), and had an open reading frame coding for 83 amino acids containing the amino acid sequence shown in Example 3(iii). The pBR322-derived plasmid containing these 308 base pairs is hereunder referred to as pHCS-1 (Example 8).

A DNA fragment containing the 308 base pairs obtained from pHCS-1 was radiolabelled by the nick translation method (see Molecular Cloning, ibid.) and, with this fragment used as a probe, the λgt10-derived cDNA library was screened by plaque hybridization [Benton and Davis, Science, 196, 180 (1977)] to obtain five clones. The nucleotde sequence of a clone which was believed to contain cDNA was determined by th same method as described above (Fig. 3).

As shown in Fig. 3, this cDNA insert had a single large open reading frame.

The amino acid sequence encoded by this cDNA can be deduced as shown in Fig. 3.

Escherichia coli strain X1776 harboring pBR322 which had this cDNA at the EcoRI cleavage site has been deposited with the Fermentation Research Institute, the Agency of Industrial Science and Technology (FERM BP-954).

This cDNA was joined to pBR327 [Soberon et al., Gene, 9, 287 (1980)] at the EcoRI site and the resulting plasmid is hereunder referred to as pBRG4.

The thus obtained pBRG4 was treated with a restriction enzyme, EcoRI, to obtain a DNA fragment containing cDNA of about 1500 base pairs. This fragment was radiolabelled by the nick translation method (see Molecular Cloning, ibid.) and, with this radiolabelled DNA fragment being used as a probe, the λgt10-derived cDNA library was screened once again by plaque hybridization (see Benton and Davis, ibid.) In this plaque hybridization, two sheets of λ-phage DNA fixed nitroc llulose filter paper were prepared; one of these sheets was used for the above-mentioned plaque hybridization and another one was subjected to plaque hybridization with th already described probe (LC). The phages which turned positive for both probes w r selected. A clone which has a "full-length" cDNA was s lected and th nucleotide sequence of

the cDNA insert as determined by the dideoxy method is shown in Fig. 4.

This cDNA had a single large open reading frame and the amino acid sequence that would be encoded by this cDNA was d duced as shown in Fig. 4 (Example 10).

Escherichia coli strain X1776 harboring pBR322 which had this cDNA at the EcoRI cleavag site has been deposited with the Fermentation Research Institute, the Agency of Industrial Science and Technology (FERM BP-955).

This cDNA was joined to pBR327 at the EcoRI site to form a plasmid which is hereunder referred to as pBRV2.

(4) Screening a human chromosomal gene library

A human chromosomal gene library that was prepared in accordance with the procedures described by Maniatis et al. (Molecular Cloning, ibid.) was subjected to screening with the pHCS-1 shown above. Probes that may be employed in screening include: a pHCS-1-derived 308-bp DNA fragment, a pBRG4-derived ca. 1500-bp DNA fragment, a pBRV2-derived ca. 1500-bp DNA fragment, a DNA fragment of an appropriate length containing part of one or more of these DNA fragments, as well as the aforementioned oligonucleotide probes [i.e., (IWQ), (A) and (IC)]. The case of using the pHCS-1 DNA fragment is hereunder described.

This DNA fragment was radiolabelled with ^{32}P in accordance with the nick translation method [see Roop et al., Cell, 15, 431 (1978)]. With the resulting ^{32}P-labelled fragment used as a probe, the human chromosomal gene library was subjected to screening by plaque hydridization (see Benton and Davis, ibid.) so as to obtain ten-odd clones.

After recovering DNA from the clones, a restriction enzyme map was prepared by known procedures [Fritsch et al., Cell, 19, 959 (1980)].

With the same DNA probe being used, Southern blotting (see Southern, ibid.) was conducted and it was found that a DNA fragment of about 4 kb that was cut out with EcoRI and XhoI could potentially contain a region for encoding the human G-CSF polypeptide. Therefore, the ca. 4-kb DNA fragment was inserted into pBR327 at the EcoRI site using an EcoRI linker so as to obtain pBRCE3β. With this plasmid being used as a base sequencing DNA, the nucleotide sequence of the ca. 3-kb portion of that ca. 4-kb DNA fragment was determined by the dideoxy method. As a result, said DNA fragment was found to be a gene coding for the human G-CSF polypeptide (Fig. 5).

E. coli strain X1776 harboring pBRCE3β (i.e. the plasmid pBR327 having said ca. 4-kb DNA fragment inserted into the EcoRI site) has been deposited with the Fermentation Research Institute, the Agency of Industrial Science and Technology (FERM BP-956).

Comparison between the pBRG4 cDNA insert shown in Fig. 3 and the pBRV2 cDNA insert shown in Fig. 4 revealed that the DNA fragment under discussion contained five exon portions and that it coded for the amino acid sequences deduced from pBRG4 and pBRV2.

Fig. 6 shows the restriction enzyme cleavage sites of the obtained gene.

This DNA fragment contained the chromosomal gene of human G-CSF, or the preceding region to be transcribed to human G-CSF mRNA, plus a nucleotide sequence taking part in transcriptional control [Benoist and Chambon, Nature, 290, 304 (1981); and Breathnack and Chambon, Ann. Rev. Biochem., 50, 349 (1981)].

(5) Construction of recombinant vectors containing the Chromosomal gene

An expression vector for COS cells was constructed by the following procedures.

The plasmid pBRCE3β that was obtained in (4) and which contained the chromosomal gene shown in Fig. 5 was treated with EcoRI.

The pSVH*K* plasmid described by Banerji et al. in Cell, 27, 299 (1981) was treated with KpnI to remove the globin gene. The plasmid was further subjected to partial digestion with HindIII so as to remov part of the late gene of SV40. The fragments were re-joined to prepare an expression vector pML-E*.

This vector was treated with the restriction enzyme, EcoRI, and dephosphorylated with an alkaline phosphatase (Takara Shuzo Co., Ltd.) to obtain a vector DNA, which was linked to the aforementioned chromosomal DNA fragment with the aid of a T4DNA ligase (Takara Shuzo Co., Ltd.) to obtain pMLCE3α which was a recombinant vector for COS cells (Example 12). As shown in Fig. 7, this plasmid contained the enhancer of SV40 gene, the replication origin of SV40, the replication origin of pBR322 and th pBR322-derived β-lactamas g ne (Amp^r), and had the human G-CSF chromosomal gen j ined downstream from the enhancer of SV40 gene.

An expression vector for C127 cells was constructed by the following procedures. A DNA fragment containing th chromosomal CSF gene was cut out with an appropriate restriction enzyme from pMLCE3α which was the expression vector for COS cells. This fragment was joined, with a T4DNA ligase, to a DNA fragment containing the origin of bovine papilloma virus (BPV) and a DNA fragment containing the early promoter of SV40. The resulting pTNCE3α was an expression vector that had a chromosomal CSF gene linked downstream from the early promoter of SV40 and which contained a 65% portion of BPV (Example 16).

The expression vector for CHO cells had two DNA fragments linked together by a T4DNA ligase; one fragment contained the chromosomal CSF gene and the early promoter of SV40 as in the case of the expression vector for C127 cells, and the other fragment contained a pAdD26SVpA-derived dhfr gene. The resulting pD26SVCE3α was an expression vector that had the chromosomal CSF gene downstream of th SV40 promoter and, the dhfr gene downstream of the principal late promoter of adenovirus (Example 18).

(6) Expression in animal cells

A representative example using COS cells as host cells is hereunder described and, for further details, see the relevant working examples.

COS cells, which were derived from monkey CV-1 cells and which had been transformed by SV40-origin deficient mutant to express the large-size T antigen of SV40 [see Gluzman et al., Cell, 32, 175 (1981)-], were transformed by the vector pMLCE3α which was obtained in (5) and which contained the human chromosomal G-CSF gene. The supernatant of the culture of the COS cells showed the human G-CSF activity (Example 13).

The COS cells were recovered and subjected to mRNA analysis, which showed the existence of two mRNAs that corresponded to the amino acid sequences depicted in Fig. 3 and Fig. 4, respectively (Example 14).

Examples

Before the present invention is described in greater detail with reference to working examples, the following referential example is provided for the purpose of illustrating the methods of assaying the CSF activity.

Referential Example: Assaying CSF Activity

The following methods were used to determine the CSF activity (hereunder abbreviated as CSA) in the present invention.

CSA assay

(a) With human bone marrow cells:

Single-layer soft agar cultivation was conducted in accordance with the method of Bradley, T.R. and Metcalf, D. (Aust. J. Exp. Biol. Med. Sci., 44, 287-300, 1966). More specifically, 0.2 ml of a bovin fetal serum, 0.1 ml of the sample, 0.1 ml of a human bone marrow nonadherent cell suspension (1 - 2 x 10^5 nuclear cells), 0.2 ml of a modified McCoy's 5A culture solution, and 0.4 ml of a modified McCoy's 5A culture solution containing 0.75% of agar were mixed, poured into a plastic dish for tissue culture (35 $mm^{\varnothing}$), coagulated, and cultured at 37°C in 5% CO_2/95% air and at 100% humidity. Ten days later, the number of colonies formed was counted (one colony consisting of at least 50 cells) and CSA was determined with one unit being the activity required for forming one colony.

(b) With mouse bone marrow cells:

A horse serum (0.4 ml), 0.1 ml of the sample, 0.1 ml of a C3H/He (female) mouse bone marrow cell suspension (0.5 - 1 x 10^5 nuclear cells), and 0.4 ml of a modified McCoy's 5A culture solution containing 0.75% of agar w re mix d, poured into a plastic dish for tissu cultur (35 $mm^{\varnothing}$),coagulated, and cultured for 5 days at 37°C in 5% CO_2/95% air and at 100% humidity. The numb r of colonies formed was counted (one colony consisting of at least 50 cells) and CSA was determined with one unit being the activity for forming one colony.

The modified McCoy's 5A culture solution used in each of the methods (a) and (b) and the human bone marrow nonadherent cell suspension used in (a) were prepared by the following procedures.

Modified McCoy's 5A culture solution (double concentration)

Twelve grams of McCoy's 5A culture solution (Gibco), 2.55 g of MEM amino acid-vitamin medium (Nissui Seiyaku Co., Ltd.), 2.18 g of sodium bicarbonate and 50,000 units of potassium penicillin G were dissolved twice in 500 ml of distilled water and the solution was aseptically filtered through a Millipore filter (0.22 μm).

Human bone marrow nonadherent cell suspension

A bone marrow fluid obtained from a healthy person by sternal puncture was diluted 5-fold with an RPMI 1640 culture solution, plated over a Ficoll-Paque solution (Pharmacia Fine Chemicals) and centrifuged at 400 x g for 30 minutes at 25°C. The interfacial cell layer (specific gravity <1.077) was recovered. The cells were washed, adjusted to a concentration of 5 x 10^6 cells/ml with an RPMI 1640 culture solution containing 20% of bovine fetal serum, poured into a 25-cm^2 plastic flask for tissue culture, and incubated for 30 minutes in a CO_2 incubator. Nonadherent cells were recovered in the supernatant, poured into a plastic flask (25 cm^2) and incubated for 2 hours and a half. Nonadherent cells in the supernatant were collected and used in an assay.

Example 1: Establishment of CHU-2

A tumor of a patient with oral cavity cancer wherein pronounced increase was observed in the numb r of neutrophiles was transplanted into nu/nu mice. About 10 days after the transplantation, the increase in th weight of the tumor and in the number of neutrophiles was significant. Twelve days after the transplantation, the tumor was extracted aseptically, dissected into cubes of 1 - 2 mm^3 and cultured in the following manner.

Ten to fifteen cubes of the tumor were put into a 50-ml plastic centrifugal tube. After addition of 5 ml of a trypsin solution (containing 0.25% of trypsin and 0.02% of EDTA), the tube was shaken for 10 minutes in a warm bath at 37°C and the supernatant was discarded. Another 5 ml of the same trypsin solution was added and trypsin digestion was conducted under agitation for 15 minutes at 37°C. The supernatant cell suspension was recovered and stored in ice after the trypsin had been inactivated by addition of 1 ml of a bovine fetal serum.

After repeating these procedures once again, the cell suspension was recovered, combined with the previously obtained suspension, and centrifuged at 400 x g for 10 minutes to obtain a cell pellet. The pellet was washed twice with F-10 containing 10% of a bovine fetal serum and was thereafter loaded in a plastic culture flask (25 cm^2) to give a cell concentration of 5 x 10^6 cells/flask. After incubation overnight in a CO_2 incubator (5% CO_2 and 100% humidity) with an F-10 culture solution containing 10% of a bovine f tal serum, the supernatant was removed together with the nonadherent cells, and culture was continued with a fresh supply of culture solution. Six days after the start of culture, the flask became full of the cells and th culture solution was replaced by a fresh one. On the next day, the culture solution was discarded and th flask was charged with 2 ml of an anti-mouse erythrocyte antibody (Cappel) diluted 5-fold with RPMI 1640 and 2 ml of a guinea pig complement (Kyokuto Seiyaku Co., Ltd.) diluted 2.5-fold with RPMI 1640. After incubation for 20 minutes at 37°C,the culture was washed twice with F-10 containing 10% of a bovine fetal serum and the nu/nu mouse derived fibroblasts were removed. Subsequently, an F-10 culture solution containing 10% of a bovine fetal serum was added and cultivation was conducted for 2 more days. Thereafter, some of the cells were recovered and subjected to cloning by the limiting dilution method.

The resulting 11 clones were checked for their CSF activity and one clone (CHU-2) exhibited activity about 10 times as high as that of the other clones.

Example 2: Isolation of CSF

The cells established in Example 1 were grown in a completely dense population (grown to confluency) in two culture flasks (150 cm^2). The cells were recovered, suspended in 500 ml of an F-10 culture solution containing 10% of a bovine fetal serum, transferred into a glass roller bottl of 1580 cm^2 (Belco), and whirl-cultured at 0.5 rpm. When the cells w re found to hav grown in a completely dens population on the inner wall of the roller bottl , th culture solution was replaced by a s rum-free RPMI 1640. After 4-day

culture, the supernatant of the culture was recovered and cultivation was continued with F-10 containing 10% of a bovine fetal serum being added. After 3-day culture, the culture solution was again replaced by a serum-free RPMI 1640 and the supernatant of the culture was recovered 4 days later. By repeating these procedures, 500 ml of the serum-free supernatant of culture p r bottle was obtained each week. In addition, this method enabled the supernatant of culture to be recovered, with the cells maintained over a significantly prolonged period.

A batch consisting of 5,000 ml of the supernatant of the culture obtained was mixed with 0.01% of Tween 20 and concentrated about 1000 times by ultrafiltration with Hollow Fiber DC-4 and Amicon PM-10 (Amicon). The concentrate was purified by the following steps.

(i) A portion (5 ml) of the concentrated supernatant of culture was subjected to gel filtration on an Ultrogel AcA54 column (4.6 cm^{ϕ} x 90 cm^{L}; LKB) at a flow rate of ca. 50 ml/hr with 0.01 M Tris-HCl buffer (pH 7.4) containing 0.15 M NaCl and 0.01% Tween 20 (Nakai Kagaku Co., Ltd.). The column had been calibrated with bovine serum albumin (Mw; 67,000), ovoalbumin (Mw; 45,000) and cytochrome C (Mw; 12,400). After completion of the gel filtration, 0.1 ml of each of the fractions was diluted 10-fold and screened for the active fractions by the above-described method of CSA assay (b). The fractions for Ve = 400 - 700 ml were found to exhibit macrophage-dominant CSA while the fractions for Ve = 800 - 1200 ml showed granulocyte-dominant CSA. Therefore, the latter fractions were collected and concentrated to a volume of ca. 5 ml on an ultrafiltration apparatus with PM-10 (Amicon).

(ii) To the cocentrated fractions was added an aqueous solution of 0.1% trifluoroacetic acid containing 30% of n-propanol (for determination of amino acid sequence; available from Tokyo Kasei K.K.) Aft r the mixture had been left to stand in ice for about 15 minutes, the precipitate was removed by centrifugation for 10 minutes at 27,000 x g. The supernatant was adsorbed on a μ-Bondapak C18 column (8 mm x 30 cm for semipreparatory use; Waters) equilibrated with the aqueous solution containing n-propanol and trifluoroacetic acid; the column was continuously eluted with an aqueous solution of 0.1% trifluoroacetic acid which contained n-propanol having a linear concentration gradient of 30 - 60%. A high performance liquid chromatographic apparatus, Hitachi Model 685-50 (Hitachi, Ltd.), and a detector, Hitachi Model 638-41 (Hitachi, Ltd.) were employed to determine the absorptions at 220 nm and 280 nm simultaneously. After elution, 10 μl of each of the fractions was diluted 100-fold and screened for the active fractions by the above-described method of CSA assay (b). The peaks eluted with 40% n-propanol were found to have CSF activity, so they were collected, re-chromatographed under the same conditions, and assayed for CSA by the same method. Again, CSF activity was observed in the peaks at 40% n-propanol. Therefore, these peaks were collected (4 fractions = 4 ml) and freeze-dried.

(iii) The freeze-dried powder was dissolved in 200 μl of an aqueous solution of 0.1% trifluoroacetic acid containing 40% of n-propanol, and the solution was subjected to high performance liquid chromotography on TSK-G 3000SW column (Toyo Soda Manufacturing Co., Ltd.; 7.5 mm x 60 cm). Elution was conducted with the same aqueous solution at a flow rate of 0.4 ml/min and the fractions were tak n in 0.4-ml portions with a fraction collector, FRAC-100 (Pharmacia Fine Chemicals). Each of the fractions taken was checked for its CSA by the same method as described above and activity was observed in the fractions for retention times of 37 - 38 minutes (corresponding to MW of ca. 2×10^4). The active factions were recovered and purified on an analytical μ-Bondapak C18 column (4.6 mm x 30 cm). The main peaks were recovered and freeze-dried. The sample obtained was assayed by the method of CSA assay (a); it was found to have human G-CSF activity.

Example 3: Determination of Amino Acid Sequence

(i) Determination of N-terminal amino acid sequence

The sample was subjected to Edman degradation with a gas-phase sequencer (Applied Biosystems) and the resulting PTH amino acid was analyzed by routine procedures with a high performanc liquid chromatographic apparatus (Beckman Instruments) and Ultrasphere-ODS column (Beckman Instruments). The column (5 μm; 4.6 mm^{ϕ} x 250 mm^{L}) was equilibrated with a starting buffer [aq. sol. containing 15 mM sodium acetate buffer (pH 4.5) and 40% acetonitrile] and injected with the sample (as dissolved in 20 μl of the starting buffer). Separation was effected by isocratic elution with the starting buffer. The flow rate was 1.4 ml/min and the column temperature was held at 40°C. Detection of the PTH amino acid was achieved utilizing the absorptions in the UV range at 269 nm and 320 nm. standard samples of PTH amino acid (Sigma) in 2-nmol portions wer separated on the same line to determine their retention times, which were compared with thos of th sample to be t sted. As a result, the sample was found to have the following amino acid sequ nc of the 40 residues from N-terminus:

H_2N - Thr - Pro - Leu - Gly - Pro - Ala - Ser - Ser -
(10)
Leu - Pro - Gln - Ser - Phe - Leu - Leu - Lys - Cys -
(20)
Leu - Glu - Gln - Val - Arg - Lys - Ile - Gln - Gly -
(30)
Asp - Gly - Ala - Ala - Leu - Gln - Glu - Lys - Leu -
(40)
Cys - Ala - Thr - Tyr - Lys -

(ii) Degradation with bromocyan

The sample was dissolved in 70% formic acid. To the solution, 200 equivalent amounts of bromocyan that had been purified by sublimation was added. The mixture was left overnight at 37°C for reaction. The reaction product was freeze-dried and fractionated by HPLC on a TSK G3000SW column (Toyo Soda Manufacturing Co., Ltd.) to obtain four peaks. The peaks were named CN-1, CN-2, CN-3 and CN-4 in th decreasing order of the molecular weight. The first two peaks (CN-1 and CN-2) had better yields and their amino acid sequences were analyzed with an automatic gas-phase sequencer (Applied Biosystems) under the same conditions as used in (i).

As a result, CN-1 was found to be a peptide from the N-terminus of G-CSF protein, and CN-2 had the followng amino acid sequence:

Pro - Ala - Phe - Ala - Ser - Ala - Phe -
Gln - Arg - Arg - Ala - Gly - Gly - Val -
Leu - Val - Ala - Ser - His - Leu - Gln -

(iii) Digestion with trypsin

The sample was dissolved in 0.1 M Tris-HCl buffer (pH 7.4) containing 8 M urea and the solution was mixed with 0.1 M Tris-HCl buffer (pH 7.4) containing 0.1% 2-mercaptoethanol to provide a final urea concentration of 2 M. A TPCK-treated trypsin (Sigma) was added such that the sample-to-enzyme ratio was 50:1. The mixture was held for 4 hours at 25°C and, after addition of an equal amount of TPCK-treated trypsin, the mixture was held for an additional 16 hours at 25°C. Thereafter, the reaction product was subjected to high-speed reverse-phased column chromatography on C8 column (Yamamura Kagaku K.K.), with elution conducted with 0.1% TFA containing n-propanol having a linear density gradient of 5 - 60%. While several peaks were obtained by measuring the absorption at 280 nm, the main peak was analyzed for its amino acid sequence with an automatic gas-phase sequencer (Applied Biosystems) under the sam conditions as used in (i). As a result, the main peak was found to be a peptide having the following sequence which contained part of the CN-2 fragment shown in (ii):

Gln - Leu - Asp - Val - Ala - Asp - Phe - Ala - Thr -
Thr - Ile - Trp - Gln - Gln - Met - Glu - Glu - Leu -
Gly - Met - Ala - Pro - Ala - Leu - Gln - Pro - Thr -
Gln - Gly - Ala - Met - Pro - Ala - Phe - Ala - Ser -

Example 4: Preparation of DNA Probe

(i) Synthesis of probe (IWQ)

Thirty successive nucleotides (see Fig. 1) were prepared on the basis of the sequence of 10 amino acids (Ile-Trp-Gln-Gln-Met-Glu-Glu-Leu-Gly-Met) included within the amino acid sequence obtained in Example 3(iii). It will be necessary to make one comment about the notation of nucleotides shown in Fig. 1; for example, the nucleotide at 9-position from 5'-terminus is an equimolar mixture of dA and dG. The starting nucleotides were mostly dimers but mononucleotides were also used as required. A glass filter equipped column was charged with 20 mg of the starting nucleotide resin, Ap-d(G) (Yamasa Shoyu Co., Ltd.). After repeated washing with methylene chloride, the 4,4'-dimethoxytrityl group was elliminated by treatment with a solution of methylene chloride containing 3% trichloroacetic acid. Subsequently, the column was washed several times with 1 ml of methylene chloride. After the column was washed with anhydrous pyridine to displace the solvent, 20 mg of a nucleotide dimer, (DMTr)ApTp(NHR_3), (Nippon Zeon; NHR_3 = triethylammonium; DMTr = dimethoxytrityl) and 0.2 ml of pyridine were added, and the interior of the column was vacuum-dried with a vacuum pump. Subsequently, 20 mg of 2,4,6-trimethylbenzenesulfonyl-3-nitrotriazolide (MSNT of Wako Pure Chemical Industries, Ltd.) and 0.2 ml of anhydrous pyridine were added, and the interior of the column was displaced with a nitrogen gas. The nucleotide resin was condensed with the dimer by reaction for 45 minutes at room temperature, with occasional shaking. After completion of the reaction, the column was washed with pyridine and the unreacted OH groups were acetylated with a pyridine solution containing excess acetic anhydride and 4-dimethylaminopyridine. After washing the column with pyridine, the following dimers or monomers were condensed, in the order written, by repeating the above-described procedures: (DMTr)Ip(NHR_3), (DMTr)-GpGp(NHR_3), (DMTr)Ip(NHR_3), an equimolar mixture of (DMTr)CpTp(NHR_3) and (DMTr)TpTp(NHR_3), an equimolar mixture of (DMTr)ApAp(NHR_3) and (DMTr)ApGp(NHR_3), an equimolar mixture of (DMTr)ApGp-(NHR_3) and (DMTr)GpGp(NHR_3), (DMTr)GpAp(NHR_3), (DMTr)TpGp(NHR_3), an equimolar mixture of (DMTr)-ApAp(NHR_3) and (DMTr)GpAp(NHR_3), (DMTr)CpAp(NHR_3), an equimolar mixture of (DMTr)ApAp(NHR_3) and (DMTr)ApGp(NHR_3), (DMTr)GpCp(NHR_3), (DMTr)TpGp(NHR_3), (DMTr)Ip(NHR_3) and (DMTr)ApTp-(NHR_3), with all of these nucleotides being available from Nippon Zeon except for (DMTr)Ip(NHR_3) which was available from Yamasa Shoyu Co., Ltd. After completion of the reaction in the final stage, the resin was washed successively with pyridine, methylene chloride and ether without acetylation, and thereafter dried. The dried resin was suspended in 1.7 ml of a mixture of pyridine (0.5 ml), water (0.2 ml) and dioxan (1 ml) containing 1 M tetramethylguanidine and 1 M α-picolinaldoxime. The suspension was left to stand overnight at room temperature and concentrated to 100 - 200 µl under vacuum. The concentrate was mixed with a small amount (2 - 3 drops) of pyridine and 2 - 3 ml of concentrated aqueous ammonia, and the mixture was heated at 55°C for 6 hours. Following extraction with ethyl acetate, the aqueous layer was separated and concentrated under vacuum. The concentrate was dissolved in a solution of 50 mM triethyl ammonium acetate (pH 7.0) and the solution was subjected to chromatography on C-18 column (1.0 x 15 cm; Waters), with elution conducted with acetonitrile (linear density gradient of 10 - 30%) in a solution of 50 mM triethyl ammonium acetate (pH 7.0). The peak fraction eluted at an acetonitrile concentration of about 25% was concentrated under vacuum.

To the concentrate, 80% acetic acid was added and the mixture was left to stand for 30 minutes at room temperature. Following extraction with ethyl acetate, the aqueous layer was separated and concentrated under vacuum. The resulting concentrate was further purified by high performance liquid chromatography on C-18 column (from Senshu Kagaku K.K.; SSC-ODS-272; 6$^{\phi}$ mm x 200 mm). Elution was conducted with acetonitrile (10 - 20% linear density gradient) in a solution of 50 mM triethyl ammonium acetate (pH 7.0). A synthetic DNA was obtained in a yield no lower than $10A_{260}$ units.

Analysis by the Maxam-Gilbert sequencing method [Meth. Enzym., 65, 499 (1980)] revealed that the oligonucleotide obtained had the nucleotide sequence shown in Fig. 1.

(ii) Synthesis of probe (A)

Fourteen successive nucleotides (see Fig. 1) were obtained on the basis of the sequence of 5 amino acids (Met-Pro-Ala-Phe-Ala) included within the amino acid sequence obtained in Example 3(iii).

Synthesis procedures were similar to those employed in the preparation of probe (IWQ), and the following nucleotides were condensed to a nucleotide resin, Ap-d(T) (Yamasa Shoyu Co., Ltd.) in the order written: (DMTr)CpAp(NHR_3), (DMTr)GpGp(NHR_3), an equimolar mixture of (DMTr)CpAp(NHR_3), (DMTr)-CpTp(NHR_3), (DMTr)CpGp(NHR_3) and (DMTr)CpCp(NHR_3), an equimolar mixture of (DMTr)ApGp(NHR_3), (DMTr)TpGp(NHR_3), (DMTr)GpGp(NHR_3) and (DMTr)CpGp(NHR_3), (DMTr)ApAp(NHR_3), an equimolar mixture of (DMTr)CpAp(NHR_3) and (DMTr)CpGp(NHR_3), and (DMTr)Gp(NHR_3), with all nucleotides being available from Nippon Zeon. A synthetic DNA was obtained in a yield of ca. $10A_{260}$ units. Analysis by the Maxam-Gilbert sequencing method revealed that the oligonucleotide obtained had the nucleotide sequence

shown in Fig. 1.

(iii) Synthesis of probe (LC)

Automatic DNA synthesis was accomplished with a DNA synthesizer, Model 380A of Applied Biosystems. This technique, based on the principles described by Caruthers et al. [J. Am. Chem. Soc., 103, 3185 (1981)], is generally referred to as the phosphoramidite procedure.

A phosphoramidite form of (DMTr)-dT preliminarily activated with tetrazole was condensed to dG-S (S: support) wherein 5'-dimethoxytrityl group (DMTr) was deblocked. Thereafter, the unreacted hydroxyl groups were acetylated and oxidated with iodine in the presence of water to make a phosphoryl group. After deblocking the DMTr group, condensation was repeated in the same manner until 24 nucleotides having the sequence shown in Fig. 1 were synthesized. These nucleotides were cleaved from the support, deblocked, and purified by reverse-phased high performance liquid chromatography on C-18 column (Senshu Kagaku Co., Ltd.; SSC-ODS-272).

Example 5: Cultivation of CHU-2 Cells and Preparation of mRNA

1) Cultivation and recovery of CHU-2 cells

Established CHU-2 cells were grown in a completely dense population (grown to confluency) in two culture flasks (150 cm^2), recovered, suspended in 500 ml of an RPMI 1640 culture solution containing 10% of a bovine fetal serum, transferred into a glass roller bottle of 1580 cm^2 (Belco), and whirl-cultured for 4 days at 0.5 rpm. When the cells were found to have grown in a completely dense population (grown to confluency) on the inner wall of the roller bottle, the culture solution was removed from the roller bottle, which was charged with 100 ml of a preheated (37°C) physiological saline solution containing 0.02% of EDTA. After heating at 37°C for 2 minutes, the cells were separated from the inner wall of the flask by pipetting. The resulting cell suspension was centrifuged at 400 x g for 10 minutes to obtain a cell pellet. The cells were resuspended in 5 ml of an EDTA-free physiological saline solution. The suspension was centrifuged at 400 x g for 10 minutes to obtain a cell pellet (wet weight, ca. 0.8 g). The so obtained cells were stored frozen at -80°C until they were subjected to procedures for extraction of RNA.

2) Purification of mRNA

Isolation of mRNA from the CHU-2 cells obtained in 1) was accomplished by procedures which were essentially the same as those described in "Molecular Cloning", Maniatis et al., Cold Spring Harbor, page 196, 1982. The frozen CHU-2 cells (wet weight, 3.8 g) were suspended in 20 ml of a solution of 6 M guanidine [6 M guanidinium isothiocyanate, 5 mM sodium citrate (pH 7.0), 0.1 M β-mercaptoethanol, and 0.5% sodium sarcosyl sulfate] and the suspension was well mixed by vortexing for 2 - 3 minutes. The mixture was subjected to 10 cyclic suction and ejection with a syringe (capacity, 20 ml) equipped with a 18G needle. About 6 ml of the viscous guanidinium solution containing the disrupted cells was layered onto a 6-ml cushion of 5.7 M CsCl in 0.1 M EDTA (pH 7.5) in a Beckman SW40 Ti polyallomer centrifuge tube in such a manner that the tube became full of the contents. Four centrifuge tubes were prepared by th procedures described above and centrifuged at 120,000 x g for 15 hours at 20°C. The resulting pellets were washed three times with a small amount of 70% ethanol.

The pellets obtained from the respective tubes were combined, dissolved in 550 µl of water and worked up to provide a NaCl concentration of 0.2 M. After treatment with a 1:1 mixture of phenol and chloroform and with chloroform alone, 2.5 volumes of ethanol were added to precipitate the total RNA (ca. 10.1 mg of the total RNA was obtained from 3.8 g of wet cells).

Poly(A^+) RNA was purified from the total RNA by the following procedures of affinity chromatography taking advantage of the attachment of a poly(A) chain at 3' terminus of the mRNA. Adsorption on oligo(dT)-cellulose (Type 7 of P-L Biochemicals) was achieved by passage through an oligo(dT)-cellulose column of the total RNA in a loading buffer [containing 10 mM Tris-HCl (pH 7.5), 0.5 M NaCl, 1 mM EDTA, and 0.1% SDS solution] after the solution had been heated at 65°C for 5 minutes. The column had been equilibrated with the same loading buffer. Elution of poly(A^+) RNA was accomplished with a TE solution [containing 10 mM Tris-HCl (pH 7.5) and 1 mM EDTA]. The unadsorbed effluent was re-charged through the column and the eluate obtained by repeating the same procedures was mixed with the first run of eluate. As a result, 400 µg of the poly(A^+) RNA was obtained.

The so prepared mRNA was fractionated for size by sucrose density gradient centrifugation in

accordanc with th procedures described in the laboratory manual of Schleif and Wensink, "Practical Methods in Molecular Biology", Springer-Verlag, New York, Heidelb rg, Berlin (1981).

Stated mor specifically, a 5 - 25% sucrose density gradient was created in a Beckman SW40 Ti centrifuge tube. Two sucrose solutions were prepared by dissolving 5% and 25% of RNase-free sucros (Schwarz/Mann) in a solution containing 0.1 M NaCl, 10 mM Tris-HCl (pH 7.5), 1 mM EDTA, arid 0.5% SDS.

Eight hundred micrograms of the mRNA [poly(A^+)-RNA] prepared by the method already described was dissolved in 200 - 500 µl of a TE solution. The solution was heated at 65°C for 5 minutes and, after being quenched, it was placed on the sucrose density gradient solutions, which were centrifuged at 120,000 x g for 20 hours. Fractions each weighing 0.5 ml were collected and their absorption at 260 nm was measured. The sizes of the fractionated RNAs were determined on the basis of the positions of standard RNAs (ribosome RNAs 282, 18S and 5S). At the same time, the G-CSF activity of each fraction was examin d with oocytes of Xenopus laevis by the following procedures. First, the mRNA of each fraction was worked up into an aqueous solution having a concentration of 1 µg/µl; oocytes were taken from Xenopus (about one year old) and the mRNA solution was injected in such a manner that a 50-ng of mRNA was injected into one oocyte; ten such oocytes were placed in each of 96 wells in a microtiter plate; the oocytes were cultured for 48 hours at room temperature in 100 µl of a Barth medium [88 mM NaCl; 1 mM KCl; 2.4 mM $NaHCO_3$; 0.82 mM $MgSO_4$; 0.33 mM $Ca(NO_3)_2$; 0.41 mM $CaCl_2$; 7.5 mM Tris-HCl (pH 7.6); penicillin, 10 mg/L; and streptomycin sulfate, 10 mg/L]; the supernatant of the culture was recovered, concentrated and purified to a grade suitable for assay of G-CSF activity.

The G-CSF activity was found to be present in 15 - 17S fractions.

Example 6: Synthesis of cDNA (Construction of pBR-line cDNA Library)

From the poly(A^+) RNA obtained in Example 5 was synthesized cDNA by the method of Land et al. [Nucleic Acids Res., 9, 2251 (1981)] as modified by the method of Gubler and Hoffman [Gene, 25, 263 (1983)].

1) Synthesis of single-stranded cDNA

An Eppendorf tube (capacity, 1.5 ml) was charged with reagents in the following order: 80 µl of a reaction buffer (500 mM KCl, 50 mM $MgCl_2$, 250 mM Tris-HCl, pH 8.3); 20 µl of 200 mM dithiothreitol, 32 µl of 12.5 mM dNTP (containing 12.5 mM each of dATP, dGTP, dCTP and dTTP), 10 µl of α-^{32}PdCTP (PB 10205 of Amerscham), 32 µl of oligo$(dT)_{12-18}$ (from P-L Biochemicals; 500 µg/ml), 20 µl of poly(A^+) RNA (2.1 µg/µl), and 208 µl of distilled water. A total of 400 µl of the reaction solution was heated at 65°C for 5 minutes, and thereafter heated at 42°C for 5 minutes. To the heated solution, 120 units of a revers transcriptase (Takara Shuzo Co., Ltd.) was added. Following reaction for 2 more hours at 42°C, 2 µl of an RNase inhibitor (Bethesda Research Laboratories), 20 µl of a TE solution, 16 µl of 100 mM sodium pyrophosphate and 48 units (4 µl) of a reverse transcriptase were added, and reaction was carried out at 46°C for 2 hours. The reaction was quenched by addition of 0.5 M EDTA (8 µl) and 10% SDS (8 µl). By subsequent treatment with phenol/chloroform and precipitation with ethanol (twice), a single-stranded cDNA was obtained.

2) Attachment of dC-chain to the single-stranded cDNA

The single-stranded cDNA obtained in 1) was dissolved in distilled water. To the solution was added 60 µl of a dC-chain adding buffer [400 mM potassium cacodylate, 50 mM Tris-HCl (pH 6.9), 4 mM dithiothreitol, 1 mM $CoCl_2$, and 1 mM dCTP], and the mixture was heated at 37°C for 5 minutes. To the reaction solution, 3 µl of a terminal transferase (27 units/µl; P-L Biochemicals) was added and the mixture was heated at 37°C for 2.5 minutes. Following treatment with phenol/chloroform (once) and precipitation with ethanol (twice), the dC-tailed cDNA was dissolved in 40 µl of a TE solution containing 100 mM NaCl.

3) Synthesis of double-stranded cDNA

To 40 µl of the DNA solution prepared in 2), 4 µl of oligo$(dG)_{12-18}$ (200 µg/ml; P-L Biochemicals) was added and th mixture was heated first at 65°C for 5 minutes, th n at 42°C for 30 minutes. While the reaction solution was h ld at 0°C, 80 µl of a buffer [100 mM Tris-HCl (pH 7.5), 20 mM $MgCl_2$, 50 mM $(NH_4)_2SO_4$, and 500 mM KCl], 4 µl of 4 mM dNTP (containing 4 mM each of dATP, dCTP, dGTP and dTTP), 60 µl of 1 mM β-NAD, 210 µl of distilled water, 20 µl of E. coli DNA polymeras I (Takara Shuzo

Co., Ltd.), 15 µl of E. coli DNA ligase (Takara Shuzo Co., Ltd.) and 15 µl of E. coli RNase H (Takara Shuzo Co., Ltd.) were added, and th mixture was subjected to reaction at 12°C for 1 hour. Following addition of 4 mM dNTP (4 µl), reaction was carried out at 25°C for 1 hour. By subsequent treatment with phenolchloroform and precipitation with ethanol (once), about 8 µg of a double-stranded cDNA was obtained. This double-stranded cDNA was dissolved in a TE solution and subjected to 1.2% agarose gel electrophoresis. Fragments corresponding to the size of ca. 560 bp to 2 kbp were adsorbed on Whatman DE81 and about 0.2 µg of the double-stranded cDNA could be recovered by elution.

4) Attachment of dC-chain to the double-stranded cDNA

The double-stranded cDNA prepared in 3) was dissolved in 40 µl of a TE solution. After 8 µl of a dC-tail adding buffer of the type identified in 2) had been added, the mixture was heated at 37°C for 2 minutes. Following addition of 1 µl of a terminal transferase (27 units/µl), the mixture was subjected to reaction at 37°C for 3 minutes. Thereafter, the reaction solution was immediately cooled to 0°C, and the reaction was quenched by addition of 1 µl of 0.5 M EDTA. Following treatment with phenol/chloroform and precipitation with ethanol, the precipitate obtained was suspended in 10 µl of a TE solution.

5) Construction of pBR-line cDNA library

Four microliters of a commercial oligo(dG)-tailed pBR322 vector (Bethesda Research Laboratories; 10 ng/µl) and 2 µl of the dC-tailed double-stranded cDNA obtained in 4) were annealed in a TE solution containing 75 µl of 0.1 M NaCl. The annealing consisted of three stages: heating at 65°C for 5 minutes, subsequent heating at 40°C for 2 hours, followed by cooling to room temperature.

In accordance with the method described in the laboratory manual of Maniatis et al. [Molecular Cloning, Cold Spring Harbor, p 249 ff. (1982)] (other routine techniques could also be used here), competent cells were prepared from E. coli strain X1776, and transformed with the annealed plasmid to produce transformants.

Example 7: Synthesis of cDNA (Construction of λphage Library)

1) Synthesis of single-stranded cDNA

In accordance with the procedures described in Example 5, 3.8 g of frozen CHU-2 cells were purified twice on an oligo(dT)-cellulose column and subsequently worked up to obtain 400 µg of poly(A^+) RNA.

A TE solution (10 µl) having 12 µg of the poly(A^+) RNA dissolved therein was placed in a reaction tube containing 10 µg of actinomycin D (Sigma). Thereafter, the tube was charged with reagents in the following order: 20 µl of a reverse transcription buffer [250 mM Tris-HCl (pH 8.3); 40 mM $MgCl_2$; 250 mM KCl]; 20 µl of 5 mM dNTP (containing 5 mM each of dATP, dGTP, dCTP and dTTP); 20 µl of oligo$(dT)_{12-18}$ (0.2 µg/ml; P-L Biochemicals); 1 µl of 1 M dithiothreitol; 2 µl of RNasin (30 units/µl; Promega Biotech); 10 µl of a reverse transcriptase (10 units/µl; Seikagaku Kogyo Co., Ltd.); 1 µl of α-^{32}P-dATP (10 µCi; Amerscham); and 16 µl of water. The reaction solution totalling a volume of 100 µl was held at 42°C for 2 hours and the reaction was quenched by addition of 0.5 M EDTA (5 µl) and 20% SDS (1 µl). By subsequent treatment with phenol/chloroform (100 µl) and precipitation with ethanol (twice), about 4 µg of a single-stranded cDNA was obtained.

2) Synthesis of double-stranded cDNA

The cDNA obtained in 1) was dissolved in 29 µl of a TE solution and a reaction solution was prepar d by adding the following reagents in the order written: 25 µl of a polymerase buffer [400 mM Hepes (pH 7.6); 16 mM $MgCl_2$, 63 mM β-mercaptoethanol, and 270 mM KCl]; 10 µl of 5 mM dNTP; 1.0 µl of 15 mM β-NAD; 1.0 µl of α-^{32}-P-dATP (10 µCi/µl); 0.2 µl of E. coli DNA ligase (60 units/µl; Takara Shuzo Co., Ltd.); 5.0 µl of E. coli DNA polymerase I (New England Biolabs; 10 units/µl); 0.1 µl of RNase H (60 units/µl; Takara Shuzo Co., Ltd.); and 28.7 µl of distilled water.

The reaction solution was incubated at 14°C for 1 hour, allowed to return to room temperature, and incubated for an additional hour. Then, th reaction was quenched by addition of 0.5 M EDTA (5 µl) and 20% SDS (1 µl), and treatm nt with ph nol/chlor form and precipitation with ethan l were performed. The DNA obtained was dissolved in 20 µl of 0.5 mM EDTA and a reaction solution was prepared by addition of 3 µl of a Klenow buffer [500 mM Tris-HCl (pH 8.0) and 50 mM $MgCl_2$], 3 µl of 5 mM dNTP, and 4 µl of

water. After addition of 1 µl of a DNA polymerase (Klenow fragment; Takara Shuzo Co., Ltd.), the reaction solution was incubated at 30°C for 15 minutes.

The incubated reaction solution was diluted with 70 µl of a TE solution and the reaction was quenched by addition of 0.5 M EDTA (5 µl) and 20% SDS (1 µl). By subsequent treatment with phenol/chloroform and precipitation with ethanol, about 8 µg of a double-stranded cDNA was obtained.

3) Methylation of double-stranded cDNA

An aqueous solution (30 µl) of the double-stranded cDNA synthesized in 2) was mixed with 40 µl of a methylation buffer [500 mM Tris-HCl (pH 8.0); 50 mM EDTA], 20 µl of a SAM solution [800 µM S-adenosyl-L-methylmethionine (SAM); 50 mM β-mercaptoethanol], and 100 µl of water. To the mixture, 15 µl of an EcoRI methylase (New England Biolabs; 20 units/µl) was added to make a reaction solution totalling 200 µl in volume. Following incubation at 37°C for 2 hours, treatments with phenol and ether and precipitation with ethanol were conducted to recover the DNA.

4) Addition of EcoRI linker

To about 1.2 µg of the methylated double-stranded DNA, 1.5 µl of a ligase buffer [250 mM Tris-HCl (pH 7.5) and 100 mM $MgCl_2$], 0.5 µl of a preliminarily phosphorylated EcoRI linker (10mer; Takara Shuzo Co., Ltd.), 1.5 µl of 10 mM ATP, 1.5 µl of 100 mM dithiothreitol, and 2 µl of H_2O were added to make a reaction solution totalling 15 µl in volume. After 0.7 µl of T_4 DNA ligase (3.4 units/µl; Takara Shuzo Co., Ltd.) had been added, reaction was carried out overnight at 4°C. Thereafter, the ligase was inactivated by heating at 65°C for 10 minutes. The reaction solution was worked up to a total volume of 50 µl by addition of 100 mM Tris-HCl (pH 7.5), 5 mM $MgCl_2$, 50 mM NaCl and 100 µg/ml of gelatin. Following addition of EcoRI (3.5 µl; 10 units/µl), reaction was carried out at 37°C for 2 hours. Subsequently, 2.5 µl of 0.5 M EDTA and 0.5 µl of 20% SDS were added, followed by treatment with phenol/chloroform and precipitation with ethanol so as to recover the DNA. Thereafter, the unreacted EcoRI linker was removed by gel filtration on Ultrogel AcA34 (LKB) or agarose-gel electrophoresis, so as to recover about 0.5 - 0.7 µg of the linker-added double-stranded cDNA.

5) Joining double-stranded cDNA to λgt10 vector

The linker-added double-stranded cDNA was mixed with 2.4 µg of preliminarily EcoRI-treated λgt10 vector (Vector Cloning system), 1.4 µl of a ligase buffer (250 mM Tris-HCl and 100 mM $MgCl_2$), and 6.5 µl of distilled water, and the mixture was heated at 42°C for 15 minutes. Thereafter, 1 µl of 10 mM ATP, 1 µl of 0.1 M dithiothreitol and 0.5 µl of T_4 DNA ligase were added to make a total volume of 15 µl and reaction was carried out overnight at 12°C.

6) In vitro packaging

About a third of the recombinant DNAs prepared in 5) was packed with an in vitro packaging kit (Promega Biotech) to obtain phage plaques.

Example 8: Screening of pBR-Line Library with Probe (IWQ)

Whatman 541 paper was placed on a colony-growing agar medium and left to stand at 37°C for 2 hours. The filter paper was subsequently treated by the following method of Taub and Thompson [Anal. Biochem., 126, 222 (1982)].

The colonies transferred onto the 541 paper were further grown onto an agar medium containing chloramphenicol (250 µg/µl) overnight at 37°C.

The 541 paper was recovered and left at room temperature for 3 minutes on another sheet of filter paper that had been impregnated with a 0.5 N NaOH solution. This procedure was repeated twice. Two similar runs were conducted for 3 minutes using a solution of 0.5 M Tris-HCl (pH 8). At 4°C, treatments were conducted with a solution of 0.05 M Tris-HCl (pH 8) for 3 minutes, and with 1.5 mg/ml of a lysozyme solution [containing 0.05 M Tris-HCl (pH 8) and 25% sucrose] for 10 minutes; then, at 37°C, treatments were conducted with a solution of 1 x SSC (0.15 M NaCl and 0.015 M sodium citrate) for 2 minutes, and with a 1 x SSC solution containing 200 µg/ml of proteinase K for 30 minutes; finally, at room temperature, treatments were conducted with a 1 x SSC solution for 2 minutes, and with 95% ethanol solution for 2

minutes. The final step was repeated twice. Thereafter, the 541 paper was dried. Th dried 541 paper was immersed in a 25:24:1 mixture of phenol/chloroform/isoamylalcohol [equilibrated with 100 mM Tris-HCl (pH 8.5), 100 mM NaCl and 10 mM EDTA] for 30 minutes at room temperatur . Subsequently, similar procedures were repeated thre times with a 5 x SSC solution for 3 minutes, then twic with a 95% ethanol solution for 3 minutes. Thereafter, the filter paper was dried.

The probe (IWQ) was labelled with ^{32}P in accordance with routine procedures (see Molecular Cloning) and colony hybridization was performed in accordance with the method of Wallace et al. [Nucleic Acids Res., 9, 879 (1981)]. Prehybridization was conducted at 65°C for 4 hours in a hybridization buffer containing 6 x NET [0.9 M NaCl; 0.09 M Tris-HCl (pH 7.5); and 6 mM EDTA], 5 x Denhardt's solution, 0.1% SDS and 0.1 mg/ml of denatured DNA (calf thymus). Thereafter, hybridization was conducted overnight at 56°C in a hybridization buffer (for its formulation, see above) containing 1 x 10^6 cpm/ml of the radiolabelled probe (IWQ). After completion of the reaction, the 541 paper was washed twice with a 6 x SSC solution (containing 0.1% SDS) for 30 minutes at room temperature, then washed at 56°C for 1.5 minutes. The washed 541 paper was then subjected to autoradiography.

The plasmid was separated from positive clones and subjected to Southern blotting with the probe (IWQ). Hybridization and autoradiography were conducted under the same conditions as described above.

Similarly, Southern blotting was conducted with the probe (A). Using a hybridization buffer having the formulation shown above, hybridisation was conducted first at 49°C for 1 hour. After leaving it to 39°C, hybridisation was further continued at the same temperature for 1 hour. After completion of the reaction, a nitrocellulose filter was washed twice with 6 x SSC containing 0.1% SDS for 30 minutes at room temperature, then washed at 39°C for 3 minutes. The washed paper was subjected to autoradiography.

As a result, a single clone was found to be positive. Nucleotide sequencing by the dideoxy method revealed that this clone had a DNA composed of 308 base pairs containing the portions of both probe (IWQ) and probe (A) (Fig. 2). The pBR322-derived plasmid containing this insert was named pHCS-1.

Example 9: Screening of λPhage Line Library with pHCS-1 Derived DNA Probe

Plaque hybridization was conducted in accordance with the method of Benton and Davis [Science, 196, 180 (1977)]. The pHCS-1 obtained in Example 8 was treated with Sau3A and EcoRI to obtain a DNA fragment of ca. 600 bp. This DNA fragment was radiolabelled by nick translation in accordance with routin procedures. A nitrocellulose filter (S & S) was placed on the phage plaque-growing agar medium to transfer the phages onto the filter. After denaturing the phage DNA with 0.5 M NaOH, the filter paper was treated by the following procedures: treatment with 0.1 M NaOH and 1.5 M NaCl for 20 seconds; two treatments with 0.5 M Tris-HCl (pH 7.5) and 1.5 M NaCl for 20 seconds; finally, treatment with 120 mM NaCl, 15 mM sodium citrate, 13 mM KH_2PO_4 and 1 mM EDTA (pH 7.2) for 20 seconds.

The filter was subsequently dried and heated at 80°C for 2 hours to immobilize the DNA. Prehybridization was conducted overnight at 42°C in a prehybridization buffer containing 5 x SSC, 5 x Denhardt's solution, 50 mM phosphate buffer, 50% formamide, 0.25 mg/ml of denatured DNA (salmon sperm DNA) and 0.1% SDS. Thereafter, hybridization was conducted at 42°C for 20 hours in a hybridization buffer containing 4 x 10^5 cpm/ml of pHCS-1 probe that had been radiolabelled by nick translation. This hybridization buffer was a mixture of 5 x SSC, 5 x Denhardt's solution, 20 mM phosphate buffer (pH 6.0), 50% formamide, 0.1% SDS, 10% dextran sulfate and 0.1 mg/ml of denatured DNA (salmon sperm DNA).

The hybridized nitrocellulose filter was washed for 20 minutes with 2 x SSC containing 0.1% SDS at room temperature, then for 30 minutes with 0.1 x SSC containing 0.1% SDS at 44°C, and finally for 10 minutes with 0.1 x SSC at room temperature. Detection by autoradiography was then conducted.

As a result, five positive clones (G1 - G5) were obtained. The clone containing a "full-length" cDNA was checked for its DNA nucleotide sequence by the dideoxy method and the nucleotide sequence shown in Fig. 3 was identified. This cDNA was cut out of the λgt10 vector and joined to pBR327 [Soberon et al., Gene, 9, 287 (1980)] at the EcoRI site to form a plasmid which could be prepared on a large scale. This plasmid is named pBRG4.

Example 10: Screening of λPhage Line Library with pBRG4-Derived DNA Probe and Probe (LC)

Plaque hybridization was performed in accordance with the method of Benton and Davis (see Science, ibid.) employed in Example 9. A nitrocellulose filter (S & S) was placed on the phage plaque-growing agar medium to transfer the phages onto the filter. After denaturing th phage DNA with 0.5 M NaOH, the filter was treated by the following procedures: treatment with 0.1 M NaOH and 1.5 M NaCl for 20 seconds; then two treatments with 0.5 M Tris-HCl (pH 7.5) and 1.5 M NaCl for 20 seconds; finally, treatment with 120 mM

NaCl, 15 mM sodium citrate, 13 mM KH_2PO_4 and 1 mM EDTA (pH 7.2) for 20 seconds. Th filter was subsequently dried, and heated at 80°C for 2 hours to immobilize the DNA. Two sheets of the same filter were prepared in th manner described above and subjected to scr ening with th pBRG4-d rived DNA probe and the probe (LC).

Screening with the pBRG4-derived DNA probe was carried out by the following procedures. The pBRG4 was treated with EcoRI to obtain a DNA fragment of ca. 1500 bp. This DNA fragment was radiolabelled by nick translation in accordance with routine procedures. One of the two nitrocellulose filters was subjected to prehybridization overnight at 42°C in a prehybridization buffer containing 5 x SSC, 5 x Denhardt's solution, 50 mM phosphate buffer, 50% formamide, 0.25 mg/ml of denatured DNA (salmon sperm DNA) and 0.1% SDS. Thereafter, the filter was subjected to hybridization at 42°C for 20 hours in a hybridization buffer containing the radiolabelled DNA probe (ca. 1 x 10^6 cpm/ml) of ca. 1500 bp. This hybridization buffer was a mixture of 5 x SSC, 5 x Denhardt's solution, 20 mM phosphate buffer (pH 6.0), 50% formamide, 0.1% SDS, 10% dextran sulfate and 0.1 mg/ml of denatured DNA (salmon sperm DNA). The hybridized nitrocellulose filter was washed for 20 minutes with 2 x SSC containing 0.1% SDS at room temperature, then for 30 minutes with 0.1 x SSC containing 0.1% SDS at 44°C, and finally for 10 minutes with 0.1 x SSC at room temperature. Detection by autoradiography was then conducted.

Screening with the probe (LC) was carried out by the following procedures. The other filter was preliminarily treated with 3 x SSC containing 0.1% SDS at 65°C for 2 hours. Then, prehybridization was conducted at 65°C for 2 hours in a solution containing 6 x NET, 1 x Denhardt's solution, and 100 µg/ml of denatured DNA (salmon sperm DNA). Hybridization was subsequently conducted overnight at 63°C in a hybridization buffer containing the radiolabelled probe (LC) (2 x 10^6 cpm/ml). This hybridization buffer was also a mixture of 6 x NET, 1 x Denhardt's solution and 100 µg/ml of denatured DNA (salmon sperm DNA). The hybridized nitrocellulose filter was washed three times (20 minutes each) with 6 x SSC containing 0.1% SDS at room temperature, then washed with 6 x SSC containing 0.1% SDS at 63° C for 2 minutes.

The filter was dried and detection was conducted by autoradiography.

In the screening described above, clones which were positive to both probes were selected and th clone containing a "full-length" cDNA was checked for its nucleotide sequence by the dideoxy method. It was found to have the nucleotide sequence shown in Fig. 4. This cDNA was cut out of the λgt10 vector and joined to pBR327 at the EcoRI site to prepare a plasmid pBRV2.

Example 11: Screening of Human Chromosomal Gene Library

1) Construction of human chromosomal gene library

The human chromosomal gene library which was provided by courtesy of Dr. Maniatis of Harvard University had been prepared by the following procedures: the whole chromosomal DNA was extracted from the human fetal liver with phenol or other appropriate chemicals and partially digested with restriction enzymes, HaeIII and AluI; the resulting DNA fragments were treated by sucrose density gradient centrifugation to concentrate the fragments having chain lengths of about 18 - 25 kb; the concentrated fragments were joined to the arm DNA of E. coli phage λ Charon 4A, with short-chained synthetic nucleotides having the cleavage sites of the restriction enzyme EcoRI being inserted, so as to prepare infectious phage DNA recombinants; with a view to providing enhanced infectiousness, more refined phage λ particles w re created by packaging. The so prepared human gene library is theoretically considered to be a set of recombinants containing human DNAs with chain lengths of 18 - 25 kb which contained practically all human genes.

2) Screening of human chromosomal gene library with the pHCS-1 derived DNA probe

Plaque hybridization was conducted in accordance with the method of Benton and Davis [Science, 196, 180 (1977)]. The pHCS-1 obtained in Example 8 was treated with Sau3A and EcoRI to obtain a DNA fragment of ca. 600 bp. This DNA fragment was radiolabelled by nick translation in accordance with routine procedures. A nitrocellulose filter (S & S) was placed on the phage plaque-growing agar medium to transfer the phages onto the filter. After denaturing the phage DNA with 0.5 M NaOH, the filter paper was treated by the following procedures: treatment with 0.1 M NaOH and 1.5 M NaCl for 20 seconds; two treatments with 0.5 M Tris-HCl (pH 7.5) and 1.5 M NaCl for 20 seconds; finally, treatment with 120 mM NaCl, 15 mM sodium citrate, 13 mM KH_2PO_4 and 1 mM EDTA (pH 7.2) for 20 seconds.

The filter was subsequently dried and heated at 80°C for 2 hours to immobilize the DNA. Prehybridization was conducted overnight at 42°C in a prehybridization buffer containing 5 x SSC, 5 x Denhardt's

solution, 50 mM phosphate buffer, 50% formamide, 0.25 mg/ml of denatured DNA (salmon sperm DNA) and 0.1% SDS. Thereafter, hybridization was conducted at 42°C for 20 hours in a hybridization buffer containing 4 x 10^5 cpm/ml of pHCS-1 probe that had been radiolabelled by nick translation. This hybridization buffer was a mixture of 5 x SSC, 5 x Denhardt's solution, 20 mM phosphat buffer (pH 6.0), 50% formamid , 0.1% SDS, 10% dextran sulfate and 0.1 mg/ml of denatured DNA (salmon sperm DNA).

The hybridized nitrocellulose filter was washed for 20 minutes with 2 x SSC containing 0.1% SDS at room temperature, then for 30 minutes with 0.1 x SSC containing 0.1% SDS at 44°C, and finally for 10 minutes with 0.1 x SSC at room temperature. Detection by autoradiography was then conducted.

As a result, ten-odd positive clones were obtained. Recombinant DNAs were prepared from these clones by the method of Maniatis [Cell, 15, 687 (1978)]. The obtained DNAs were treated with restriction enzymes such as EcoRI, BamHI and BglII, analyzed by agarose gel electrophoresis, and their restriction enzyme map was prepared in accordance with the method of Fritsch et al. (see Cell, ibid.)

Southern hybridization was conducted with the probe being the radiolabelled pHCS-1 derived DNA fragment that was the same as what was used in the above-described screening procedures. A DNA fragment of ca. 8 kb that was cut with EcoRI was selected from the clones that hybridized with the probe. This fragment was subcloned to the EcoRI site of pBR327. The subcloned DNA was subjected to another treatment with restriction enzymes and Southern hybridization was conducted repeatedly. A DNA fragment of ca. 4 kb that was cut out with EcoRI and XhoI was found to contain a gene coding for the human G-CSF polypeptide. This DNA fragment was checked for the sequence of its ca. 3-kb portion by the dideoxy method and the nucleotide sequence shown in Fig. 5 was identified. This DNA fragment had the restriction enzyme cleavage sites shown in Fig. 6.

Screening of human chromosomal genes was also conducted using pBRG4-derived DNA and pBRV2-derived DNA as probes. In either case, a DNA fragment of 1500 bp that had been treated with EcoRI was directly radiolabelled by nick translation in the manner described above or, alternatively, a DNA fragment of ca. 700 bp that was obtained by successive treatments with EcoRI and DraI was radiolabelled by nick translation. The so prepared probe was used in plaque hybridization that was conducted under the same conditions as described above. Selected clones were analyzed by Southern hybridization so as to obtain a DNA fragment having the nucleotide sequence shown in Fig. 5. The plasmid thus obtained was named pBRCE3β.

Example 12: Construction of Recombinant Vector Containing Chromosomal Gene for Expression in COS Cells

The plasmid pBRCE3β that was obtained in Example 11 and which contained the chromosomal gene shown in Fig. 5 was treated with EcoRI. The pSVH*K* plasmid described by Banerji et al. in Cell, 27, 299 (1981) was treated with KpnI to remove the globin gene. The plasmid was further subjected to partial digestion with HindIII so as to remove part of the late gene of SV40. The fragments were rejoined to prepare an expression vector pML-E*.

This vector was treated with the restriction enzyme, EcoRI, and dephosphorylated with an alkaline phosphatase (Takara Shuzo Co., Ltd.) to obtain a vector DNA, which was linked to the aforementioned chromosomal DNA with the aid of a T4DNA ligase (Takara Shuzo Co., Ltd.) to obtain pMLCE3α. As shown in Fig. 7, this plasmid contained the enhancer of SV40 gene, the replication origin of SV40, the replication origin of pBR322 and the pBR322-derived β-lactamase gene (Amp^r), and had the human G-CSF chromosomal gene joined downstream from the enhancer of SV40 gene.

Example 13: Expression of Human G-CSF Chromosomal Gene in COS Cells

COS-1 cells (provided by courtesy of Dr. Gluzman of Cold Spring Harbor Laboratory, U.S.A.) that had been grown to a density of about 70% in Petri dishes (9 cm$^\phi$, Nunc) using a DMEM medium (Dulbecco's modified Eagle's medium available from Nissui Seiyaku K.K. under the trade name "Nissui") containing 10% calf serum were transformed by either the calcium phosphate procedure [Wigler et al., Cell, 14, 725 (1978)] or the DEAE-dextran:chloroquine method [see, for example, Gordon et al., Science, 228, 810 (1985)-].

Transformation by the calcium phosphate procedure was conducted as follows: 160 μg of the plasmid pMLCE3α prepared in Example 12 was dissolved in 320 μl of a TE solution and, after addition of distilled water (3.2 ml), 504 μl of 2 M $CaCl_2$ was added.

To the resulting solution, 4 ml of 2 x HBS (50 mM Hepes, 280 mM NaCl, 1.5 mM phosphate buffer, pH 7.12) was added and the mixture was cooled on ice for 20 - 30 minutes. Th cooled mixture was added

dropwise to the medium in an amount of 1 ml per Petri dish where the COS-1 cells had grown. After cultivation for 4 hours at 37°C in a CO_2 incubator, the cells were washed with a serum-free DMEM medium, then left to stand for about 3 minutes at room temperatur in 5 ml of a DMEM medium containing 20% glycerol, and rewashed with a serum-free DMEM medium. After the serum-free DMEM medium was removed, 10 ml of a DMEM medium containing 10% calf serum was added and cultivation was conducted overnight in a CO_2 incubator. After the medium was replaced by a fresh one of the same type, cultivation was conducted for an additional 3 days.

Transformation by the DEAE-dextran:chloroquine method was conducted as follows: as in the calcium phosphate procedure, COS-1 cells were cultivated to grow to a density of 70% and washed twice with a serum-free DMEM medium; to the washed cells, a serum-free DMEM medium containing 250 μg/ml of DEAE-dextran and 2 μg/ml of the plasmid pMLCE3α prepared in Example 12 was added and cultivation was conducted at 37°C for 12 hours; subsequently, the cells were washed twice with a serum-free DMEM medium and subjected to further cultivation at 37°C for 2 hours in a DMEM medium containing 10% calf serum and 1 mM chloroquine; thereafter, the cells were washed twice with a serum-free DMEM medium and cultured at 37°C for an additional 3 days in a DMEM medium containing 10% calf serum.

The supernatant of the so obtained culture of COS-1 cells was adjusted to a pH of 4 with 1 N acetic acid. After addition of an equal volume of n-propanol, the resulting precipitate was removed by centrifugation. The supernatant was passed through an open column (1^{ϕ} cm x 2 cm^{L}) filled with a C8 reverse-phased carrier (Yamamura Kagaku K.K.) and elution was conducted with 50% n-propanol. The eluate was diluted two-fold with water and subjected to reverse-phased high performance liquid chromatography on YMC-C8 column (Yamamura Kagaku K.K.), followed by elution with n-propanol (30 - 60% linear density gradient) containing 0.1% TFA. The fractions which were eluted at n-propanol concentrations of about 40% were recovered, freeze-dried and dissolved in 0.1 M glycidine buffer (pH 9). As a result of these procedures, the human G-CSF in the supernatant of the culture of COS-1 cells was concentrated about 20-fold.

As controls, COS-1 cells were transformed with G-CSF chromosomal-gene free pML-E^{+} by the above-described procedures and the supernatant of the resulting culture was concentrated.

The human G-CSF activities of the obtained samples were assayed by the "Method of Human G-CSF Activity Assay (a)" described earlier in this specification. The results are summarized in Table 1.

Table 1

	Human neutrophilic colonies (colonies/dish)
Purified human G-CSF (20 ng)	18
Culture of COS cells transformed with pML-E^{+} (concentrated 20-fold)	0
Culture of COS cells transformed with pMLCE3α (concentrated 20-fold)	23
Culture of COS cells transformed with pMLCE3α (concentrated 10-fold)	19

Example 14: RNA Analysis of G-CSF

COS cells cultivated to a cell concentration of 8 x 10^{6} cells/plate (9 cm^{ϕ}) were transformed with 80 μg of the plasmid pMLCE3α. After 48 hours, the totel RNA was prepared in accordance with the procedure of Chirgwin [Biochemistry, 18, 5294 - 5299 (1979)].

The plasmid pBRG4 obtained in Example 9 was cleaved with restriction enzyme AhaIII and the resulting pBRG4-derived DNA fragment was radiolabelled with [γ-^{32}P]ATP using T4 polynucleotide kinase to obtain

an ca. 2.8-kb DNA fragment containing G-CSF cDNA. The fragment was recovered and used as a DNA probe. After the DNA probe (1.5×10^5 c.p.m., 2.8×10^6 c.p.m./µg DNA) was denatured, it was mixed with 20 µg of the total RNA prepared from COS cells. Hybridization at 45°C for 15 hours was conducted. The mixture was digested with 200 units/ml or 400 units/ml of SI nuclease (P.L. Biochemicals) in accordance with the procedures of Weaver and Weissmann [Nucleic Acid Res., 7, 1175 - 1193 (1979)], followed by 4% polyacrylamide gel electrophoresis in the presence of 8.3 M urea. Detection by autoradiography was then conducted.

As a result, a band corresponding to 722 bp was observed as a strongly radiolabelled band in COS cells, from which a band corresponding to 487 bp was also detected.

Therefore, the RNA of COS cells was found to contain G-CSF mRNAs that corresponded to the amino acid sequences depicted in Fig. 3 and Fig. 4, respectively.

Example 15: Physicochemical Properties of the Obtained G-CSF

1) Analysis of amino acid composition

The crude CSF sample prepared in Example 13 was purified in accordance with the procedures described in Example 2(iii). The purified CSF sample was hydrolyzed by routine procedures, and the amino acid composition of the protein portion of the hydrolyzate was analyzed by a method of amino acid analysis with an automatic amino acid analyzer, Hitachi 835 (Hitachi Ltd.) The results are shown in Table 2. Hydrolysis was conducted under the following conditions:

(i) 6 N HCl, 110°C, 24 hours, in vacuum

(ii) 4 N methanesulfonic acid + 0.2% 3-(2-aminoethyl)indole, 110°C, 24 hours, 48 hours, 72 hours, in vacuum

The sample was dissolved in a solution (1.5 ml) containing 40% n-propanol and 0.1% trifluoroacetic acid. Aliquots each weighing 0.1 ml were dried with a dry nitrogen gas and, after addition of the reagents listed in (i) or (ii), the containers were sealed in vacuum, followed by hydrolysis of the contents.

Each of the values shown in Table 2 was the average of four measurements, 24 hour value for (i) and 24, 48 and 72 hour values for (ii), except that the contents of Thr, Ser, 1/2 Cys, Met, Val, Ile and Trp were calculated by the following methods (see "Tampaku Kagaku (Protein Chemistry) II", A Course in Biochemical Experiments, Tokyo Kagaku Dohjin):

- For Thr, Ser, 1/2 Cys and Met, the time-dependent profile of the 24, 48 and 72 hour values for (ii) was extrapolated by zero hours.
- For Val and Ile, the 72 hour value for (ii) was used.
- For Trp, the average of 24, 48 and 72 hour values for (ii) was used.

Table 2

Amino Acid Analysis Data

Amino acids	Mole%
Asp (Asp + Asn)	2.3
Thr	3.9
Ser	8.3
Glu (Glu + Gln)	15.3
Pro	7.4
Gly	7.9
Ala	10.8
1/2 Cys	2.8
Val	4.3
Met	1.7
Ile	2.3
Leu	18.7
Tyr	1.7
Phe	3.4
Lys	2.3
His	2.9
Trp	1.1
Arg	2.9

2) Sugar composition analysis

An internal standard (25 nmol of inositol) was added to 200 ng of the purified CSF sample used in th analysis of amino acid composition 1). After addition of a methanol solution (500 µl) containing 1.5 N HCl, reaction was carried out at 90°C for 4 hours in a N_2 purged, closed tube. After the tube was opened, silver carbonate (Ag_2CO_3) was added to neutralize the contents. Thereafter, 50 µl of acetic anhydride was added and the tube was shaken for an adequate period. Subsequently, the tube was left overnight in the dark at room temperature. The upper layer was put into a sample tube and dried with a nitrogen gas. Methanol was added to the precipitate and the mixture was washed and lightly centrifuged. The upper layer was put into the sam sample tube and dried. After addition of 50 µl of a TMS reagent (5:1:1 mixture of pyridine, hexamethyl disilazane and trimethylchlorosilane), reaction was carried out at 40° C for 20 minutes and the reaction product was stored in a d p freezer. A standard was prepared by combining 25 nmol of inositol with 50 nmol each of galactose (Gal), N-acetyl galactosamine (Gal NAc), sialic acid and any other

appropriate reagents.

The samples thus prepared were subjected to gas chromatographic analysis under the following conditions:

Conditions of analysis

Column :	2% OV - 17 VINport HP, 60 - 80 mesh, 3 m, glass
Temperature :	elevated from 110 to 250°C at 4°C/min.
Carrier gas (N_2) pressure :	initially 1.2 - 1.6 kg/cm^2
	finally 2 - 2.5 kg/cm^2
Sensitivity :	10 MΩ range, 0.1 - 0.4 volts
Pressure :	H_2, 0.8 kg/cm^2
	air, 0.8 kg/cm^2
Sample feed :	2.5 - 3.0 µl.

As a result of the analysis, galactose, N-acetyl galactosamine and sialic acid were identified in the CSF sample of the present invention.

3) Molecular weight

The molecular weight of the CSF sample used in the analysis of amino acid composition 1) was determined by sodium dodecylsulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The electrophoretic equipment was PROTEANTH (16 cm, product of Bio-Rad Corporation), using a gel made up of a polyacrylamide slab gel (T = 15%, C = 2.6%) measuring 140 mm x 160 mm x 1.5 mm, and a concentrating gel (T = 3%, C = 20%). A denatured CSF sample was prepared by the following procedure: CSF was boiled for 3 minutes in a solution containing 2% of sodium dodecylsulfate in 0.46 M 2-mercaptoethanol. After performing electrophoresis with 4 µg of the sample with a constant current of 30 mA for 4 hours, the gel was removed and stained with 0.25% Coomassie Brilliant Blue R 250 (product of Sigma Chemical Co.) for band detection. The following substances were used as molecular weight markers after similar treatments: phosphorylase B (mol. wt. 92,500), bovine serum albumin (BSA, mol. wt. 67,000), ovalbumin (OVA, mol. wt. 45,000), carbonic anhydrase (mol. wt. 31,000), soybean trypsin inhibitor (mol. wt. 21,500) and lysozyme (mol wt. 14,400).

As a result, a single band correponding to a molecular weight of 19,000 ± 1,000 was detected from the CSF sample.

4) Isoelectric point

The purified CSF sample used in the analysis of amino acid composition in (1) was employed.

The isoelectric point of the CSF of the present invention was determined by a flat bed, isoelectric electrophoretic apparatus, FBE-3000 (product of Pharmacia Fine Chemicals). After 2-hour electrophoresis with a constant power of 30 watts (Vmax = 2,000 volts) on a polyacrylamide gel (T = 5%, C = 3%, 115 mm x 230 mm) containing Pharmalyte (pH = 4 - 6.5, Pharmacia Fine Chemicals) and 4M urea, the CSF was fixed with 30% methanol/10% trichloroacetic acid/35% sulfosalicylic acid, and stained with Coomassie Brilliant Blue R-250. A Low pI kit (pH: 2.5 - 6.5, product of Pharmacia Fine Chemicals) was used as a isoelectric point marker.

Analysis of band separation at a pH of 4 to 6.5 gave three distinct bands corresponding to pI = 5.5, 5.8 and 6.1 for the CSF sample.

Example 16: Construction of Recombinant Vector Containing Chromosomal Gene for Expression in C127 Cells

The plasmid pMLCE3α obtained in Example 12 was treated with EcoRI and a fragment of ca. 4 kb was recovered by the procedures described in Molecular Cloning, ibid. The recovered fragment was used as a source of the chromosomal G-CSF gene.

The fragment was treated with the Klenow fragment of DNA polym rase I to create blunt ends (A).

In a separate way, the EcoRI fragment prepared in Example 9 which had the cDNA shown in Fig. 3 was treated with a restriction enzym , DraI, at 37°C for 2 hours, followed by tr atment with the Klenow fragment of DNA polymeras I (Takara Shuzo Co., Ltd.) to create blunt nds. One microgram of BglII linker (8mer, Takara Shuzo Co., Ltd.) was phosphorylated with ATP and joined to about 1 µg of th separately obtained

mixture of DNA fragments. The joined fragments were treated with a restriction enzyme, BglII, and subjected to agarose gel electrophoresis. Subsequently, only the largest DNA fragment was recovered.

This DNA fragment was equivalent to about 710 base pairs containing a human G-CSF polypeptide coding portion. A vector pdKCR [Fukunaga et al., Proc. Natl. Acad. Sci., USA, 81, 5086 (1984)] was treated with a restriction enzyme, BamHI, and subsequently dephosphorylated with an alkali phosphatase (Takara Shuzo Co., Ltd.). The vector DNA obtained was joined to the 710-bp cDNA fragment in the presence of T_4 DNA ligase (Takara Shuzo Co., Ltd.), so as to produce pHGA410.

The promoter of SV40 (ca. 0.4-kb EcoRI-EcoRI fragment) was cut out from this plasmid pHGA410 by the procedures described in Molecular Cloning, ibid., and was subsequently treated with the Klenow fragment of DNA polymerase (B).

In a separate step, a plasmid pdBPV-1 having a bovine papilloma virus (BPV) [this plasmid was obtained by courtesy of Dr. Howley and is described in Sarver, N., Sbyrne, J.C. & Howley, P.M., Proc. Natl. Acad. Sci., USA, 79, 7147-7151 (1982)] was treated with HindIII and PvuII to obtain a DNA fragment of ca. 8.4 kb. This fragment was treated with the Klenow fragment of DNA polymerase I and dephosphorylated with a bacterial alkaline phosphatase (C).

The DNA fragments (A), (B) and (C) each weighing 0.1 µg were dissolved in 20 µl of a reaction solution [50 mM Tris-HCl (pH 7.6), 10 mM $MgCl_2$, 10 mM DTT, 1 mM ATP] and reaction was carried out overnight at 4°C in the presence of 180 units of a T4DNA ligase.

The reaction solution was subsequently treated by the rubidium chloride procedure described in Molecular Cloning, ibid. so as to obtain the plasmid pTNCE3α (Fig. 8).

The resulting pTNCE3α was an expression vector that had a chromosomal CSF gene linked downstream from the early promoter of SV40 and which contained a 65% portion of BPV.

The DNA fragment (A) used as a source of the chromosomal G-CSF gene may be replaced by a DNA fragment of ca. 1.78 kb that is obtained by the following procedures: 20 µg of pMLCE3α is dissolved in 100 µl of a mixture of 10 mM Tris-HCl (pH 8.0), 7 mM $MgCl_2$, 100 mM NaCl, 7 mM 2-mercaptoethanol and 0.01% BSA; the solution is incubated at 37°C for 5 hours in the presence of 20 units of StuI and subjected to electrophoresis through 1.2% agarose gel.

Example 17: Transformation of C127 Cells and G-CSF Expression Therein

Before it was used to transform mouse C127 cells, the pTNCE3α obtained in Example 16 was treated with a restriction enzyme, BamHI. Twenty micrograms of the plasmid pTNCE3α was dissolved in 100 µl of a reaction solution [10 mM Tris-HCl (pH 8.0), 7 mM $MgCl_2$, 100 mM NaCl, 2 mM 2-mercaptoethanol and 0.01% BSA] and treated with 20 units of BamHI (Takara Shuzo Co., Ltd.), followed by treatments with phenol and ether, and precipitation with ethanol.

Mouse C127I cells were grown in a Dulbecco's minimal essential medium containing 10% bovine fetal serum (Gibco). The C127I cells growing on plates (5 cm^{ϕ}) were transformed with 10 µg, per plate, of the separately prepared DNA by the calcium phosphate procedure [see Haynes, J. & Weissmann, C., Nucleic Acids Res., 11, 687-706 (1983)]. After treatment with glycerol, the cells were incubated at 37°C for 12 hours. The incubated cells were transferred onto three fresh plates (5 cm^{ϕ}) and the media were changed twic a week. At day 16, the foci were transferred onto fresh plates and subjected to serial cultivation on a Dulbecco's minimal essential medium containing 10% bovine fetal serum (Gibco), so as to select clones having high G-CSF production rate.

Example 18: Construction of Recombinant Vector Containing Chromosomal Gene for Expression in CHO Cells

(1) Construction of the plasmid pD26SVCE3α

As in the case of expression in C127 cells, the plasmid pMLCE3α was treated with StuI and a DNA fragment of ca. 1.78 kb was recovered; alternatively, the same plasmid was treated with EcoRI and an EcoRI fragment of about 4 kb was recovered. Either fragment was suitable for use as a source of the chromosomal G-CSF gene.

The source fragment was treated with the Klenow fragment of DNA polymerase I (a).

As in Example 16, the promoter of SV40 (EcoRI-EcoRI fragment) was cut out from pHGA410 to obtain a fragment of about 0.4 kb, which was similarly treated with the Klenow fragment of DNA polymerase I (b).

In a separate step, the plasmid pAdD26SVpA [Kaufman, R.G. & Sharp, P.A., Mol. Cell. Biol., 2, 1304-1319 (1982)] was treated with EcoRI, then with the Klenow fragment of DNA polymerase, and finally

dephosphorylated by treatment with a bacterial alkalin phosphatase (c).

The fragments, (a), (b) and (c), each weighing 0.1 μg were dissolved in 20 μl of a reaction solution [50 mM Tris-HC1 (pH 7.6), 10 mM $MgCl_2$, 10 mM DTT, 1 mM ATP] and reaction was carried out overnight at 4°C in the presenc of 180 units of a T4DNA ligase.

The reaction solution was subsequently treated by the rubidium chloride preeedure described in Molecular Cloning, ibid., so as to transform E. coli strain DHl. The resulting Tet^r colonies were screened for those containing the plasmid pD26SVCE3α.

As shown in Fig. 9, the plasmid pD26SVCE3α has the CSF gene linked to the early gene of SV40, and the dhfr gene linked downstream from the principal late promoter of adenovirus.

(2) Construction of the plasmid pDRCE3α

The plasmid pAdD26SVpA was treated with EcoRI and BamHI so as to obtain a DNA fragment (ca. 2 kb) containing dhfr gene (d).

In a separate step, the plasmid pHGA410 (Example 16) was partially digested with EcoRI to create blunt ends. A linker HindIII was attached to the DNA, which was subsequently treated with HindIII and T4DNA ligase. The treated DNA was used to transform E. coli strain DHl by the rubidium chloride procedure. The resulting plasmid was named pHGA410(H). From the pHGA410(H), EcoRI - SalI fragment was prepared (e).

The fragments (d), (a) obtained in (l) and (e) were linked so as to construct an Amp^r expression vector pDRCE3α (Fig. 9).

Example 19: Transformation of CHO Cells and G-CSF Expression Therein

CHO cells were transformed with the so obtained plasmids, pD26SVCE3α and pDRCE3α by the following procedures.

CHO cells (dhfr$^-$ strain; obtained by courtesy of Dr. L. Chasin of Columbia University) were cultivated for growth in alpha-minimal essential medium containing 10% calf serum (α-MEN supplemented with adenosine, deoxyadenosine and thymidine) in plates (9 $cm^{\varnothing}$, Nunc). The cultured cells were transformed by the calcium phosphate procedure [Wigler et al., Cell, 14, 725 (1978)] in the following manner.

A carrier DNA (calf thymus DNA) was added in an appropriate amount to 1 μg of the plasmid pD26SVCE3α prepared in 1) of Example 18, and the mixture was dissolved in 375 μl of a TE solution, followed by addition of 125 μl of 1 M $CaCl_2$. After the solution was cooled on ice for 3 - 5 minutes, 500 μl of 2 x HBS (50 mM Hepes, 280 mM NaCl, and 1.5 mM phosphate buffer) was added to the solution. After re-cooling on ice, the solution was mixed with 1 ml of the culture of CHO cells, transferred onto plates, and incubated for 9 hours in a CO_2 incubator. The medium was removed from the plate and, following washing with TBS (Tris-buffered saline), addition of 20% glycerol-containing TBS, and re-washing, a non-selective medium (the α-MEN medium described above except that it was supplemented with nucleotides) was added. After 2-day incubation, a 10-fold dilution of the culture was transferred onto a selective medium (not supplemented with nucleotides). The cultivation was continued, with the medium being replaced by a fresh selective medium every 2 days, and the resulting colonies were selected and transferred onto fresh plates, where the cells grew in the presence of 0.02 μM methotrexate (MTX), followed by cloning through growth in the presence of 0.05 μM MTX, which was later increased to 0.1 μM.

The transformation of CHO cells was also accomplished with the plasmid pDRCE3α by increasing MTX concentrations so as to obtain G-CSF producing colonies.

Example 20: Assay of the G-CSF Activity of Transformants (expressing human chromosomal gene)

The supernatants of cultures of C127 cells and CHO cells which were obtained in Examples 17 and 19, respectively, were worked up as in Example 13 to obtain human G-CSF and its activity was assayed. The results are shown in Table 3.

Table 3

Assay of Human G-CSF Activity

		Human neutrophilic colonies (colonies/dish)
Purified human G-CSF (20 ng)		85
BPV	Culture of C127 cells transformed with pdBPV-1 (concentrated 20-fold)	0
	Culture of C127 cells transformed with pTNCE3α (concentrated 20-fold)	83
dhfr	Culture of CHO cells transformed with pAdD26SVpA (concentrated 20-fold)	0
	Culture of CHO cells transformed with pD26SVCE3α (concentrated 20-fold)	85
	Culture of CHO cells transformed with pDRCE3α (concentrated 20-fold)	86

Cell line CHU-2 was deposited with Institut Pasteur on September 12, 1985 and received the deposition No. I-483

Claims

1. A human chromosomal gene coding for a polypeptide having a human granulocyte colony stimulating factor activity having all or part of the nucleotide sequence shown in Figure 5.

2. A human chromosomal gene according to Claim 1 wherein said human chromosomal gene contains a nucleotide sequence that takes part in transcriptional control.

3. A human chromosomal gene according to Claim 1 which is connected to a microorganism- or virus-derived replicon.

4. A recombinant vector containing a human chromosomal gene according to claim 1.

5. A recombinant vector according to Claim 4 wh rein said human chromosomal gene contains a nucleotide sequence that takes part in transcriptional control.

6. A recombinant vector according to Claim 4 or 5 wherein said human chromosomal gene is connected to a microorganism-or virus-derived replicon.

7. A transformant containing a recombinant vector according to any one of claims 4 to 6.

8. A process for producing a glycoprotein having a human granulocyte colony stimulating factor activity which comprises:
   a) culturing the transformant according to claim 7 in a culture medium and
   b) recovering from the culture a glycoprotein having the human granulocyte colony stimulating factor activity.

9. A process according to Claim 8 wherein said glycoprotein has a sugar chain portion and a polypeptide which is represented by all or part of the following amino acid sequence:

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln
Ser Phe Leu Leu Lys Cys Leu Glu Gln Val Arg
Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu
Lys Leu (Val Ser Glu)$_m$ Cys Ala Thr Tyr Lys Leu
Cys His Pro Glu Glu Leu Val Leu Leu Gly His
Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser
Cys Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys
Leu Ser Gln Leu His Ser Gly Leu Phe Leu Tyr
Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln
Leu Asp Val Ala Asp Phe Ala Thr Thr Ile Trp
Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala
Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe
Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly Val
Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu
Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln
Pro , and

where m is 0 or 1 .

10. E. coli strain X 1776 harboring a ca. 4 kb DNA fragment coding for human G-CSF inserted into th EcoRI site of plasmid pBR327 (FERM BP-956).

Revendications

1. Gène chromosomique humain codant pour un polypeptide ayant une activité de facteur de stimulation de colonie de granulocytes humains comprenant tout ou partie de la séquence de nucléotides qu montre la Fig. 5.

2. Gène chromosomique humain suivant la revendication 1, dans lequel le gène chromosomique humain contient une séquence de nucléotides qui prend part au contrôle transcriptionnel.

3. Gène chromosomique humain suivant la revendication 1, qui st uni à un réplicon dérivant d'un micro-organism ou d'un virus.

4. Vecteur recombinant contenant un gène chromosomique humain suivant la revendication 1.

5. Vecteur recombinant suivant la revendication 4, dans lequel le gène chromosomique humain contient une séquenc de nucléotides qui prend part au contrôle transcriptionnel.

6. Vecteur recombinant suivant la revendication 4 ou 5, dans lequel le gène chromosomique humain est uni à un réplicon dérivant d'un micro-organisme ou d'un virus.

7. Transformant contenant un vecteur recombinant suivant l'une quelconque des revendications 4 à 6.

8. Procédé de production d'une glycoprotéine ayant une activité de facteur de stimulation de colonie de granulocytes humains, qui comprend :
   a) la culture du transformant suivant la revendication 7 dans un milieu de culture, et
   b) l'isolement, à partir de la culture, d'une glycoprotéine ayant l'activité de facteur de stimulation de colonie de granulocytes humains.

9. Procédé suivant la revendication 8, dans lequel la glycoprotéine comprend une partie de chaîne de sucre et une partie polypeptide qui est représentée par tout ou partie de la séquence d'acides aminés ci-après :

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln
Ser Phe Leu Leu Lys Cys Leu Glu Gln Val Arg
Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu
Lys Leu (Val Ser Glu)$_m$ Cys Ala Thr Tyr Lys Leu
Cys His Pro Glu Glu Leu Val Leu Leu Gly His
Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser
Cys Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys
Leu Ser Gln Leu His Ser Gly Leu Phe Leu Tyr
Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln
Leu Asp Val Ala Asp Phe Ala Thr Thr Ile Trp
Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala
Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe
Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly Val
Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu
Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln
Pro, et

où m est 0 ou 1.

10. Souche X 1776 d'E. coli hébergeant un fragment d'ADN d'environ 4 kilobases codant pour le G-SCF humain inséré dans le site EcoRI du plasmide pBR327 (FERM BP-956).

Patentansprüche

1. Menschliches chromosomales Gen, das ein Polypeptid mit Aktivität von menschliche Granulozyten-Kolonie-stimuli render Faktor codiert, das die gesamte oder ein n T il der in Figur 5 darg stellten Nucleotidsequenz aufweist.

2. Menschliches chromosomales Gen nach Anspruch 1, wobei das menschliche chromosomale Gen eine Nucleotidsequenz enthält, die an der transkriptionalen Kontrolle beteiligt ist.

3. Menschliches chromosomales Gen nach Anspruch 1, das mit einem von einem Mikroorganismus oder Virus abgeleiteten Replikon verbunden ist.

4. Rekombinanter Vektor, der ein menschliches chromosomales Gen nach Anspruch 1 enthält.

5. Rekombinanter Vektor nach Anspruch 4, wobei das menschliche chromosomale Gen eine Nucleotidsequenz enthält, die an der transkriptionalen Kontrolle beteiligt ist.

6. Rekombinanter Vektor nach Anspruch 4 oder 5, wobei das menschliche chromosomale Gen mit einem von einem Mikroorganismus oder Virus abgeleiteten Replikon verbunden ist.

7. Transformante, die einen rekombinanten Vektor nach einem der Ansprüche 4 bis 6 enthält.

8. Verfahren zur Herstellung eines Glykoproteins mit der Aktivität von menschliche Granulocyten-Kolonie-stimulierendem Faktor, umfassend
   a) Züchtung der Transformante nach Anspruch 7 in einem Kulturmedium und
   b) Gewinnung eines Glykoproteins mit der Aktivität von menschliche Granulocyten-Kolonie-stimulierendem Faktor aus der Kultur.

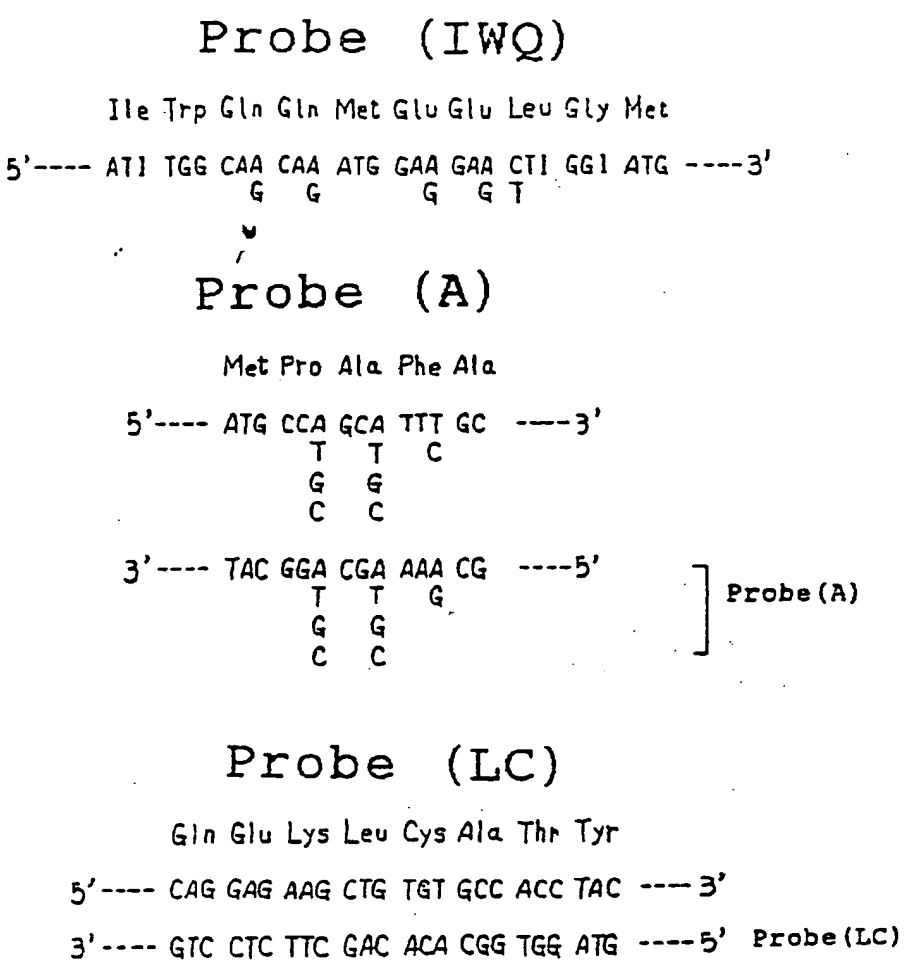
9. Verfahren nach Anspruch 8, wobei das Glykoprotein einen Zuckerketten-Anteil und ein Polypeptid enthält, das durch die gesamte oder einen Teil der folgenden Aminosäuresequenz dargestellt wird:

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln
Ser Phe Leu Leu Lys Cys Leu Glu Gln Val Arg
Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu
Lys Leu (Val Ser Glu)$_m$ Cys Ala Thr Tyr Lys Leu
Cys His Pro Glu Glu Leu Val Leu Leu Gly His
Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser
Cys Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys
Leu Ser Gln Leu His Ser Gly Leu Phe Leu Tyr
Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser
Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln
Leu Asp Val Ala Asp Phe Ala Thr Thr Ile Trp
Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala
Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe
Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly Val
Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu
Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln
Pro ,

wobei m 0 oder 1 ist.

10. E. coli Stamm X 1776, der ein m nschlichen G-CSF codierendes, etwa 4 kb großes DNA-Fragment enthält, das an der EcoRI-Stelle von Plasmid pBR327 eingefügt ist (FERM BP-956).

Fig. 1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Ile Trp Gln Gln Met Glu Glu Leu Gly Met
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G G G G T
Probe (A)
Met Pro Ala Phe Ala
5'---- ATG CCA GCA TTT GC ----3'
T T C
G G
C C
3'---- TAC GGA CGA AAA CG ----5'
T T G
G G
C C
Probe(A)
Probe (LC)
Gln Glu Lys Leu Cys Ala Thr Tyr
5'---- CAG GAG AAG CTG TGT GCC ACC TAC ---3'
3'---- GTC CTC TTC GAC ACA CGG TGG ATG ----5'
Probe(LC)

Fig. 2

CC CTG GAA GGG ATC TCC CCC GAG
TTG GGT CCC ACC TTG GAC ACA CTG
CAG CTG GAC GTC GCC GAC TTT GCC
ACC ACC ATC TGG CAG CAG ATG GAA
GAA CTG GGA ATG GCC CCT GCC CTG
CAG CCC ACC CAG GGT GCC ATG CCG
GCC TTC GCC TCT GCT TTC CAG CGC
CGG GCA GGA GGG GTC CTA GTT GCC
TCC CAT CTG CAG AGC TTC CTG GAG
GTG TCG TAC CGC GTT CTA CGC CAC
CTT GCC CAG CCC TGA GCC AAG CCC
TCC CCA TCC CAT GTA TTT ATC TCT
ATT TAA TAT TTA TGT CTA TTT

Fig. 3-1

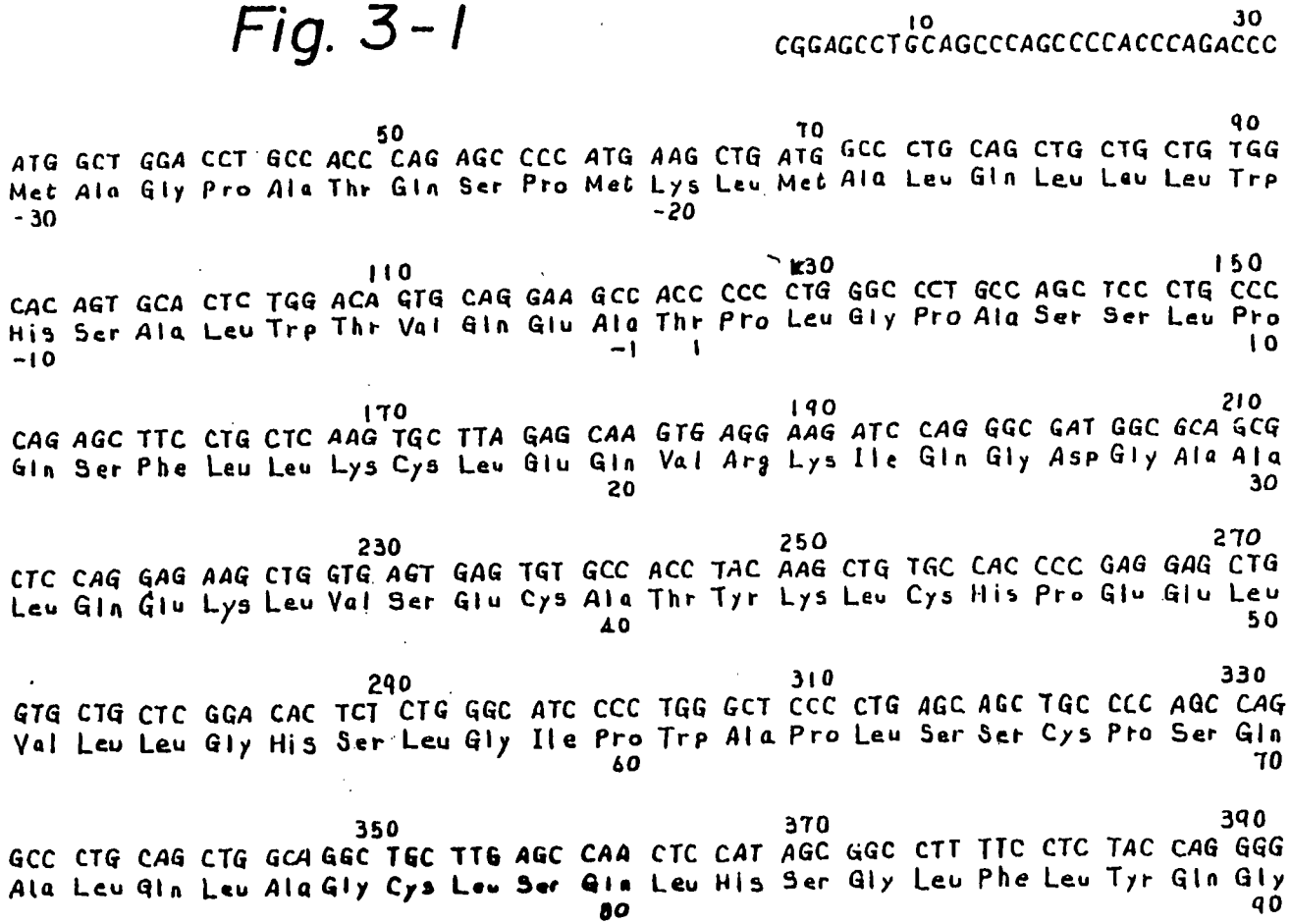

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Fig. 3-2

```
                                410                             430                             450
CTC CTG CAG GCC CTG GAA GGG ATC TCC CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG
Leu Leu Gln Ala Leu Glu Gly Ile Ser Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln
                                100                                             110

                                470                             490             510
CTG GAC GTC GCC GAC TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC
Leu Asp Val Ala Asp Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala
                                120                                             130

                                530                             550             570
CCT GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC CAG CGC CGG
Pro Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe Gln Arg Arg
                                140                                             150

                                590                             610             630
GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC CTG GAG GTG TCG TAC CGC GTT
Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu Val Ser Tyr Arg Val
                                160                                             170

                        650                             670                             690
CTA CGC CAC CTT GCC CAG CCC TGA GCCAAGCCCTCCCCATCCCATGTATTTATCTCTATTTAATATTTATG
Leu Arg His Leu Ala Gln Pro End

      710                             730                             750                             770
TCTATTTAAGCCTCATATTTAAAGACAGGGAAGAGCAGAACGGAGCCCCAGGCCTCTGTGTCCTTCCCTGCATTTCTG

      790                             810                             830                             850
AGTTTCATTCTCCTGCCTGTAGCAGTGAGAAAAAAGCTCCTGTCCTCCCATCCCCTGGACTGGGAGGTAGATAGGTAAAT

      870                             890                             910                             930
ACCAAGTATTTATTACTATGACTGCTCCCCAGCCCTGGCTCTGCAATGGGCACTGGGATGAGCCGCTGTGAGCCCCTGG
```


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Fig. 3 - 3

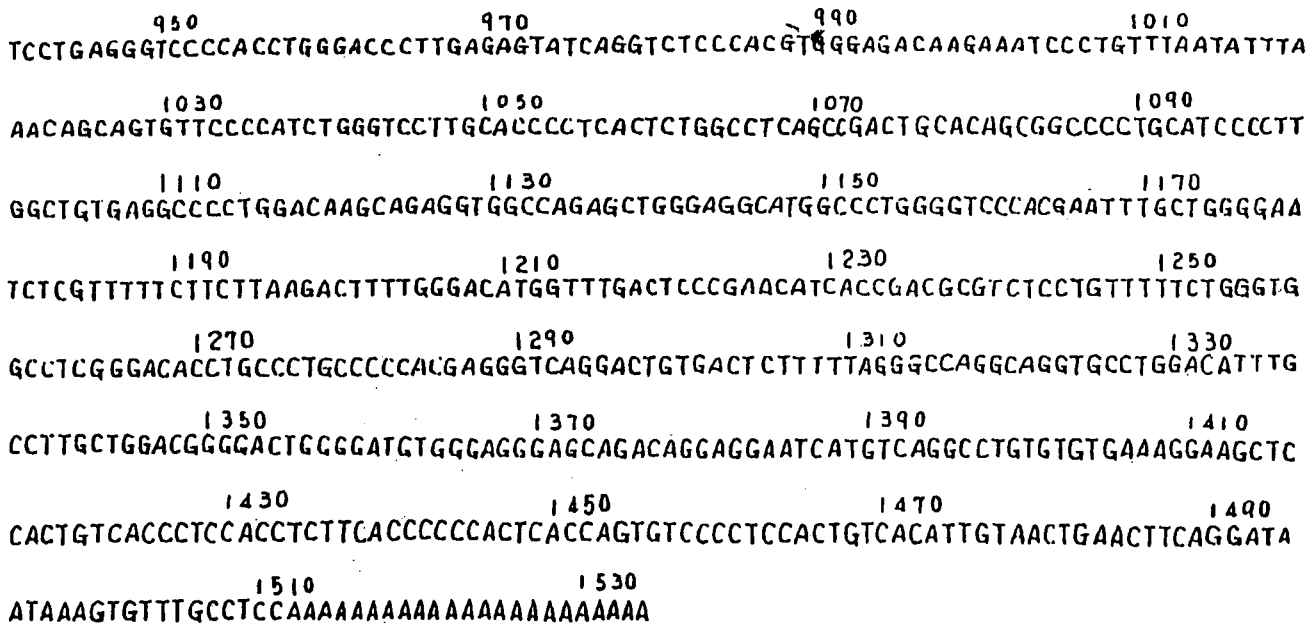

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Fig. 4-1

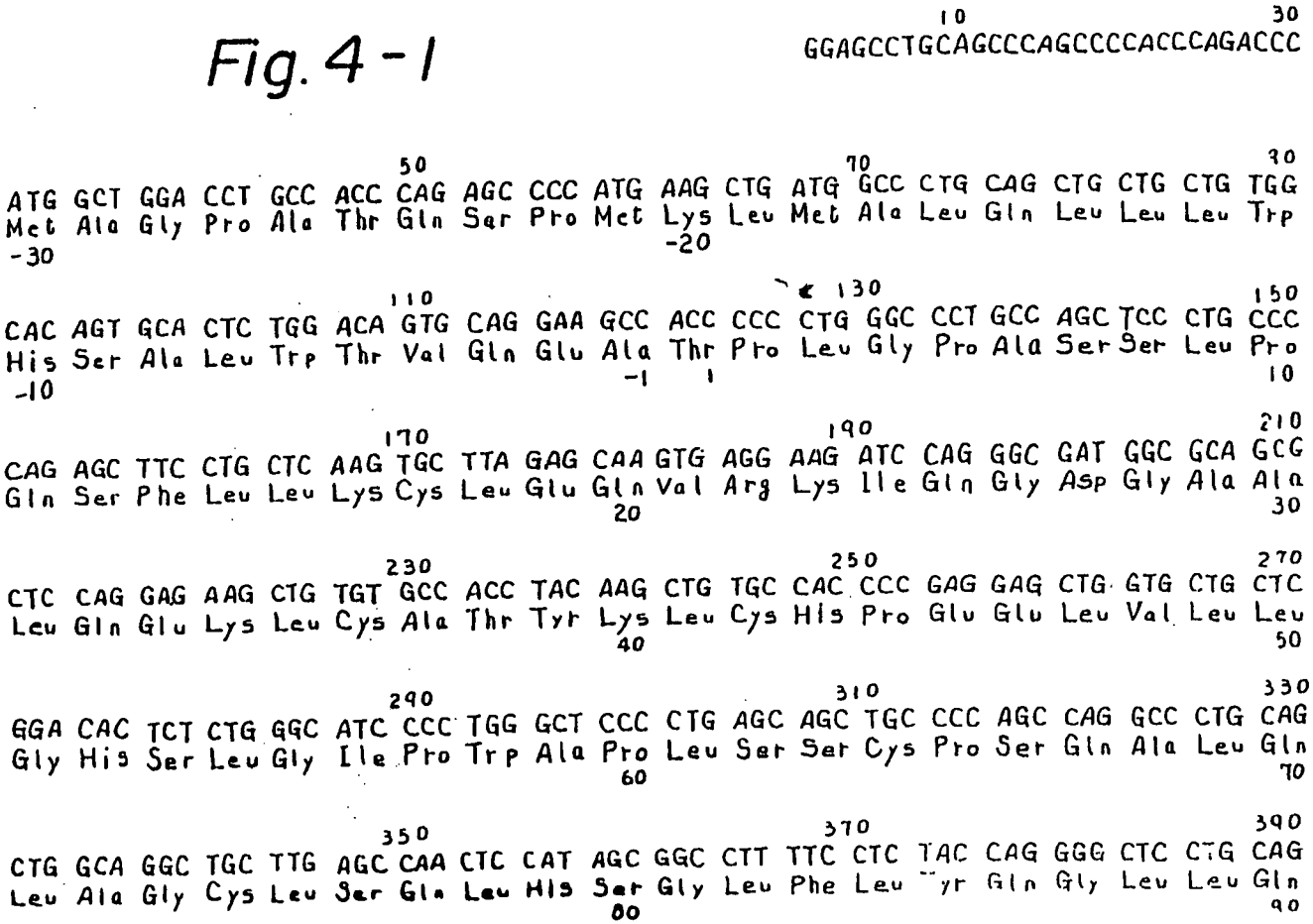

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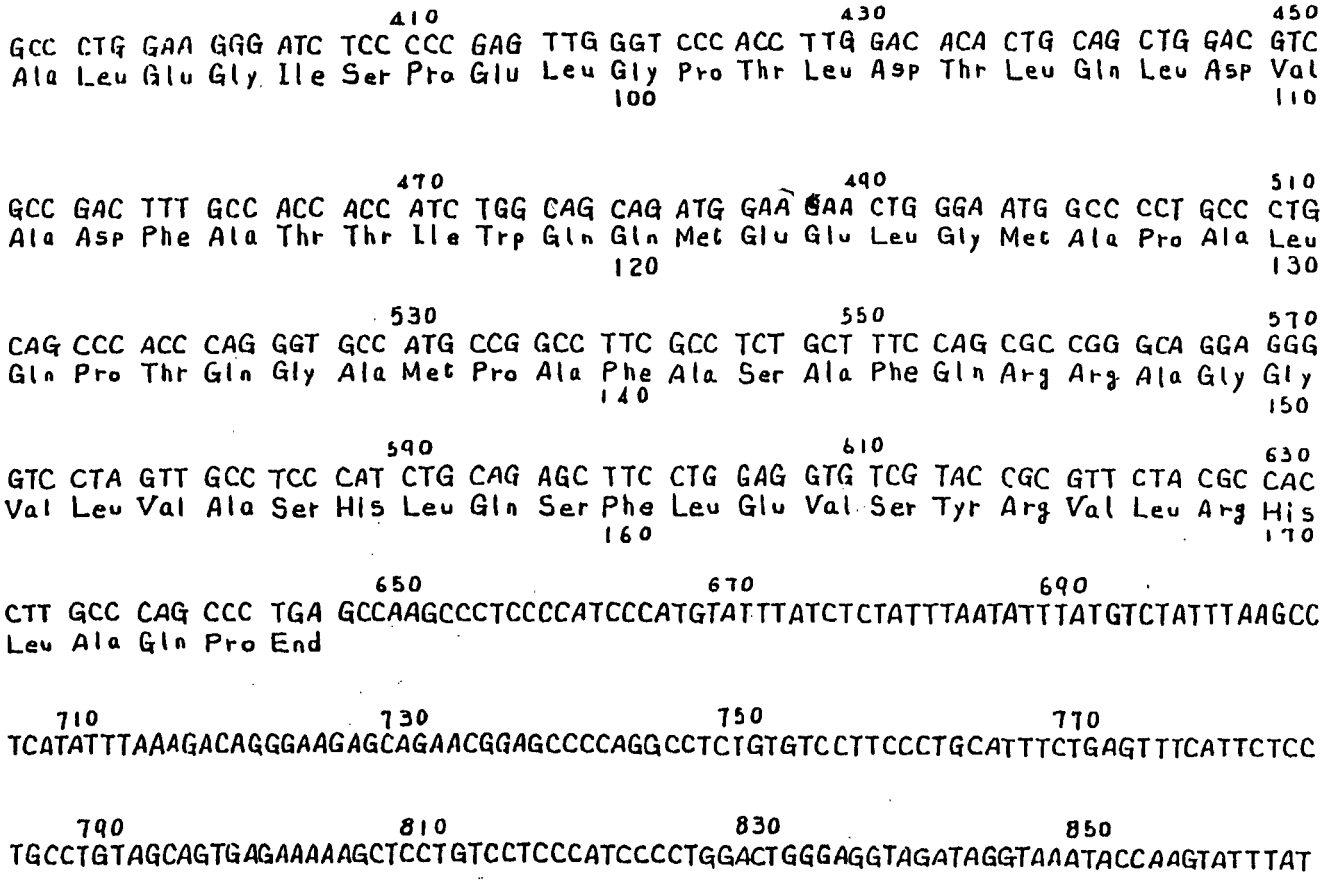
Fig. 4-2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Ala Leu Glu Gly Ile Ser Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val
100 110
470 490 510
GCC GAC TTT GCC ACC ACC ATC TGG CAG CAG ATG GAA GAA CTG GGA ATG GCC CCT GCC CTG
Ala Asp Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala Leu
120 130
530 550 570
CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC CAG CGC CGG GCA GGA GGG
Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly
140 150
590 610 630
GTC CTA GTT GCC TCC CAT CTG CAG AGC TTC CTG GAG GTG TCG TAC CGC GTT CTA CGC CAC
Val Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu Val Ser Tyr Arg Val Leu Arg His
160 170
650 670 690
CTT GCC CAG CCC TGA GCCAAGCCCTCCCCATCCCATGTATTTATCTCTATTTAATATTTATGTCTATTTAAGCC
Leu Ala Gln Pro End
710 730 750 770
TCATATTTAAAGACAGGGAAGAGCAGAACGGAGCCCCAGGCCTCTGTGTCCTTCCCTGCATTTCTGAGTTTCATTCTCC
790 810 830 850
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Fig. 4-3

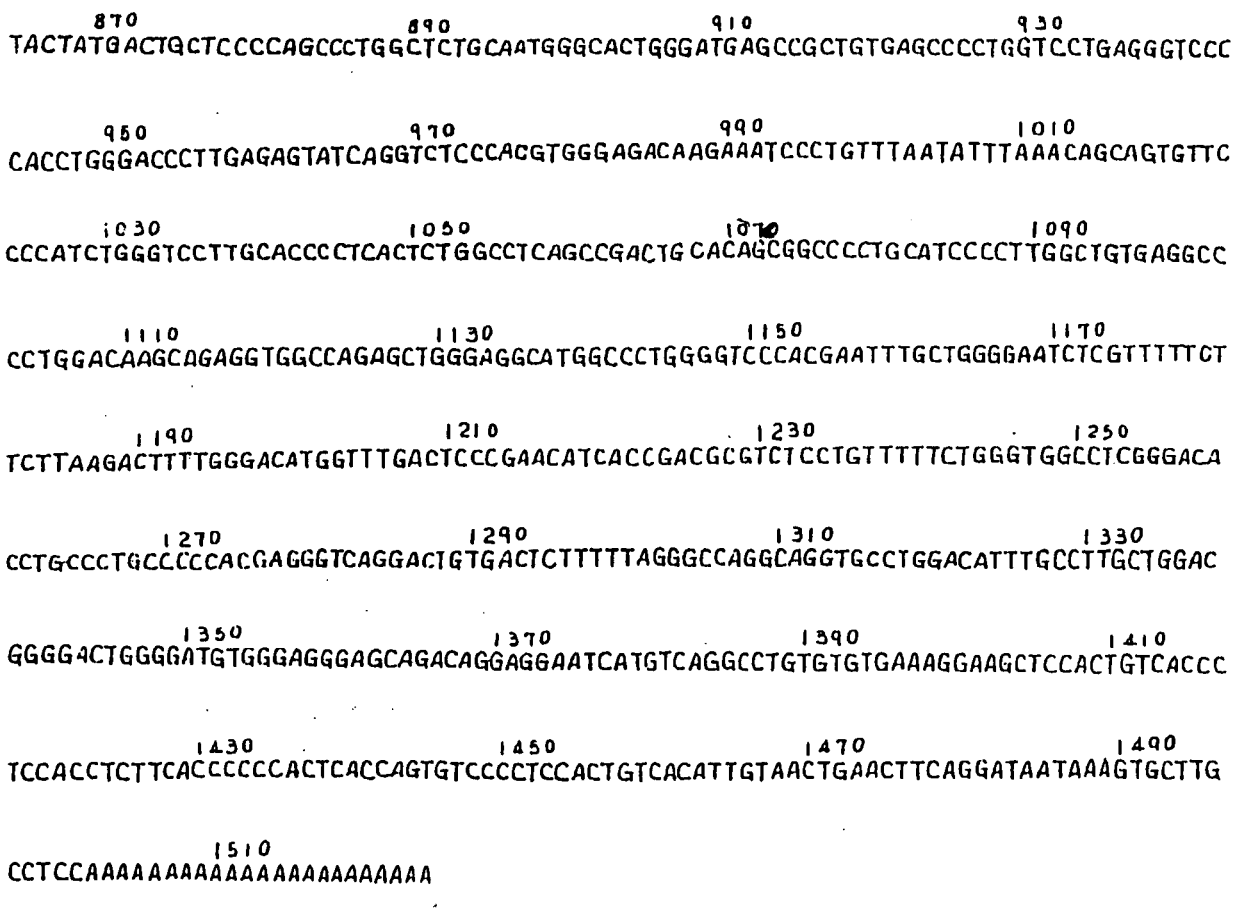

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Fig. 5 - 1

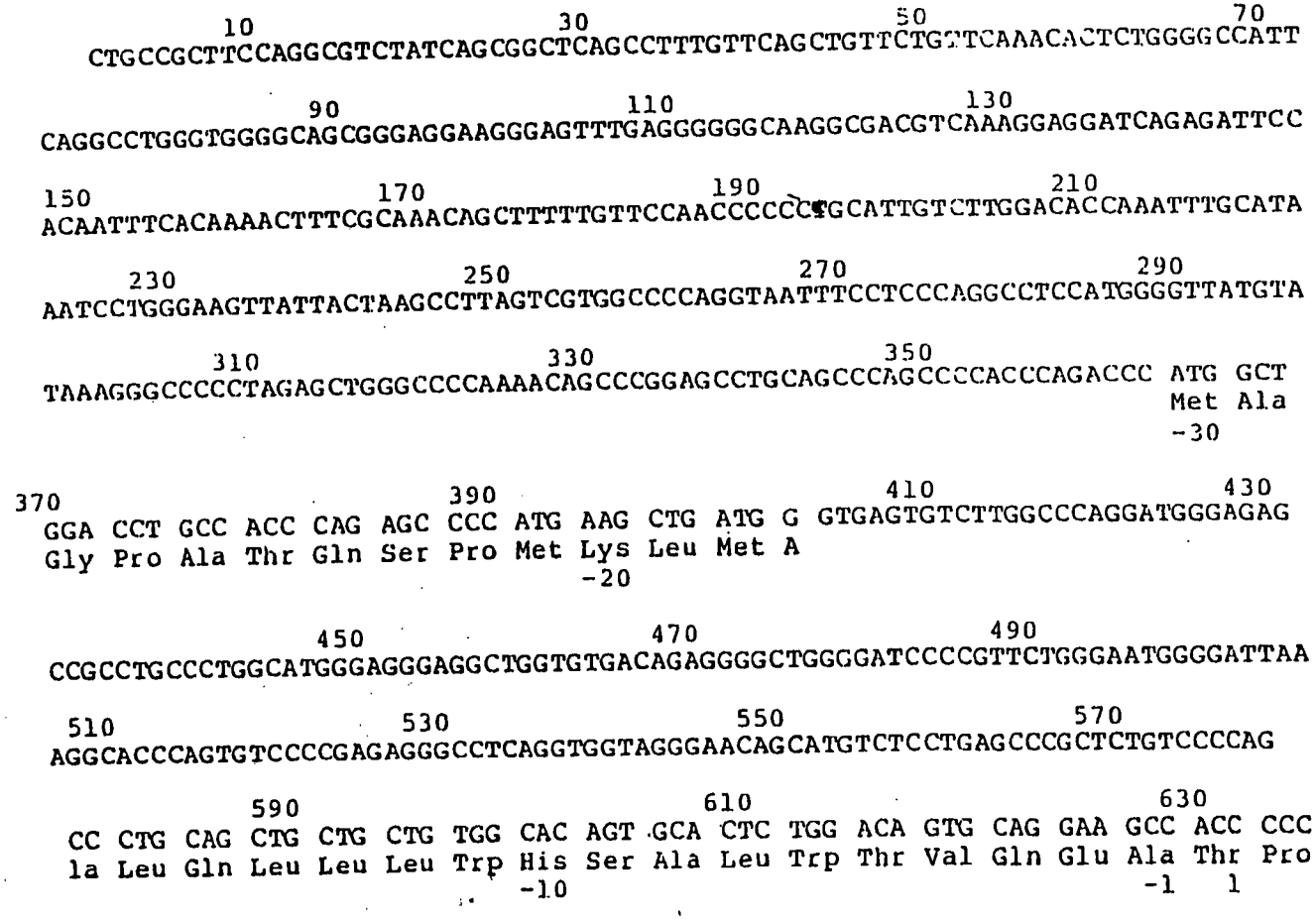

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Fig. 5-2

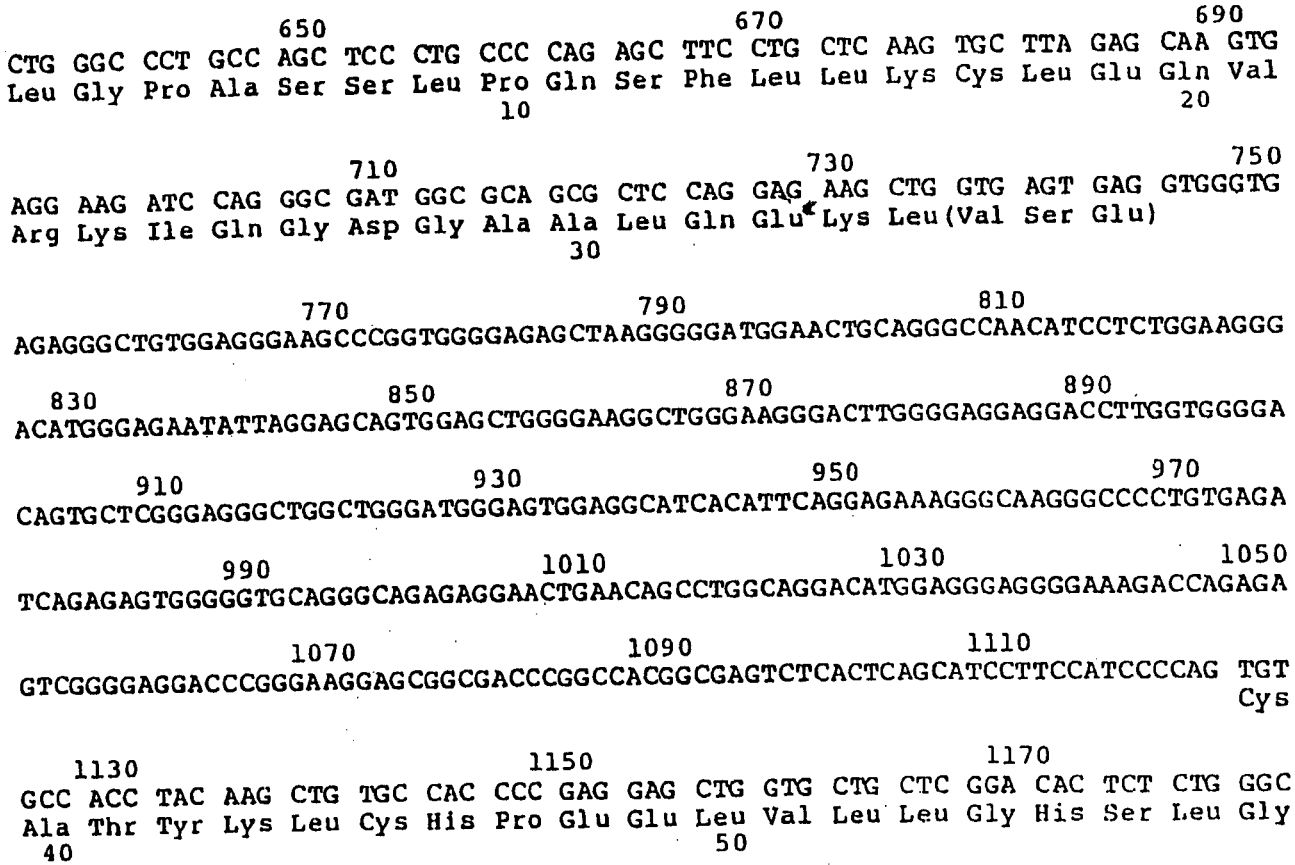

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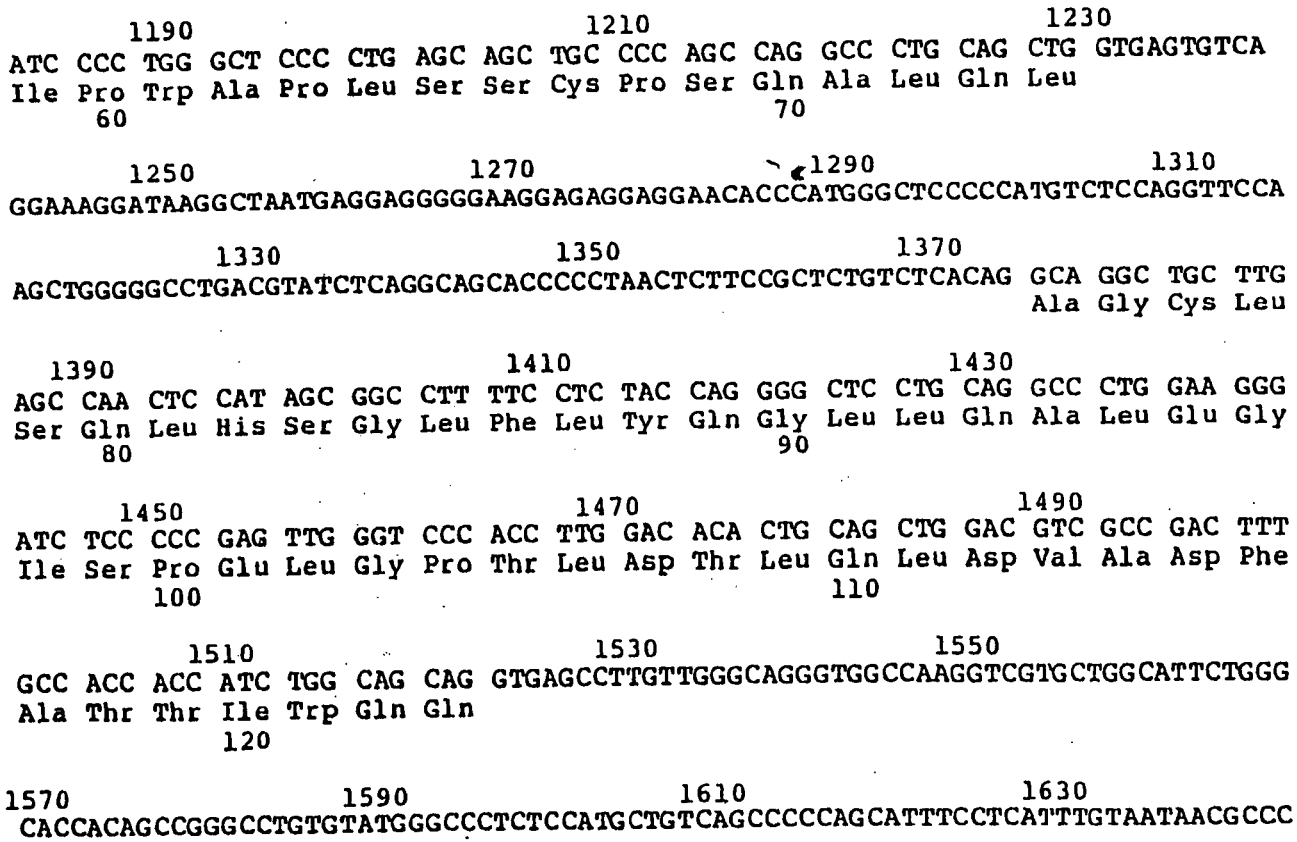
Fig. 5-3


1190
1210
1230
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Ile Pro Trp Ala Pro Leu Ser Ser Cys Pro Ser Gln Ala Leu Gln Leu
60
70
1250
1270
1290
1310
GGAAAGGATAAGGCTAATGAGGAGGGGGAAGGAGAGGAGGAACACCCATGGGCTCCCCCATGTCTCCAGGTTCCA
1330
1350
1370
AGCTGGGGGCCTGACGTATCTCAGGCAGCACCCCCTAACTCTTCCGCTCTGTCTCACAG GCA GGC TGC TTG
Ala Gly Cys Leu
1390
1410
1430
AGC CAA CTC CAT AGC GGC CTT TTC CTC TAC CAG GGG CTC CTG CAG GCC CTG GAA GGG
Ser Gln Leu His Ser Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly
80
90
1450
1470
1490
ATC TCC CCC GAG TTG GGT CCC ACC TTG GAC ACA CTG CAG CTG GAC GTC GCC GAC TTT
Ile Ser Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp Phe
100
110
1510
1530
1550
GCC ACC ACC ATC TGG CAG CAG GTGAGCCTTGTTGGGCAGGGTGGCCAAGGTCGTGCTGGCATTCTGGG
Ala Thr Thr Ile Trp Gln Gln
120
1570
1590
1610
1630
CACCACAGCCGGGCCTGTGTATGGGCCCTCTCCATGCTGTCAGCCCCCAGCATTTCCTCATTTGTAATAACGCCC

Fig. 5-4

```
    1650                1670                1690
ACTCAGAAGGGCCCAACCACTGATCACAGCTTTCCCCCACAG ATG GAA GAA CTG GGA ATG GCC CCT
                                            Met Glu Glu Leu Gly Met Ala Pro
                                                                       130

1710                1730                1750
GCC CTG CAG CCC ACC CAG GGT GCC ATG CCG GCC TTC GCC TCT GCT TTC CAG CGC CGG
Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe Gln Arg Arg
                            140                                        150

  1770                1790                1810
GCA GGA GGG GTC CTG GTT GCC TCC CAT CTG CAG AGC TTC CTG GAG GTG TCG TAC CGC
Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu Val Ser Tyr Arg
                                    160

      1830                1850                1870
GTT CTA CGC CAC CTT GCC CAG CCC TGA GCCAAGCCCTCCCCATCCCATGTATTTATCTCTATTTAA
Val Leu Arg His Leu Ala Gln Pro End
170

1890                1910                1930                1950
TATTTATGTCTATTTAAGCCTCATATTTAAAGACAGGGAAGAGCAGAACGGAGCCCCAGGCCTCTGTGTCCTTCC

    1970                1990                2010                2030
CTGCATTTCTGAGTTTCATTCTCCTGCCTGTAGCAGTGAGAAAAAGCTCCTGTCCTCCCATCCCCTGGACTGGGA

          2050                2070                2090                2110
GGTAGATAGGTAAATACCAAGTATTTATTACTATGACTGCTCCCCAGCCCTGGCTCTGCAATGGGCACTGGGATG

                2130                2150                2170
AGCCGCTGTGAGCCCCTGGTCCTGAGGGTCCCCACCTGGGACCCTTGAGAGTATCAGGTCTCCCACGTGGGAGAC
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Fig. 5-5

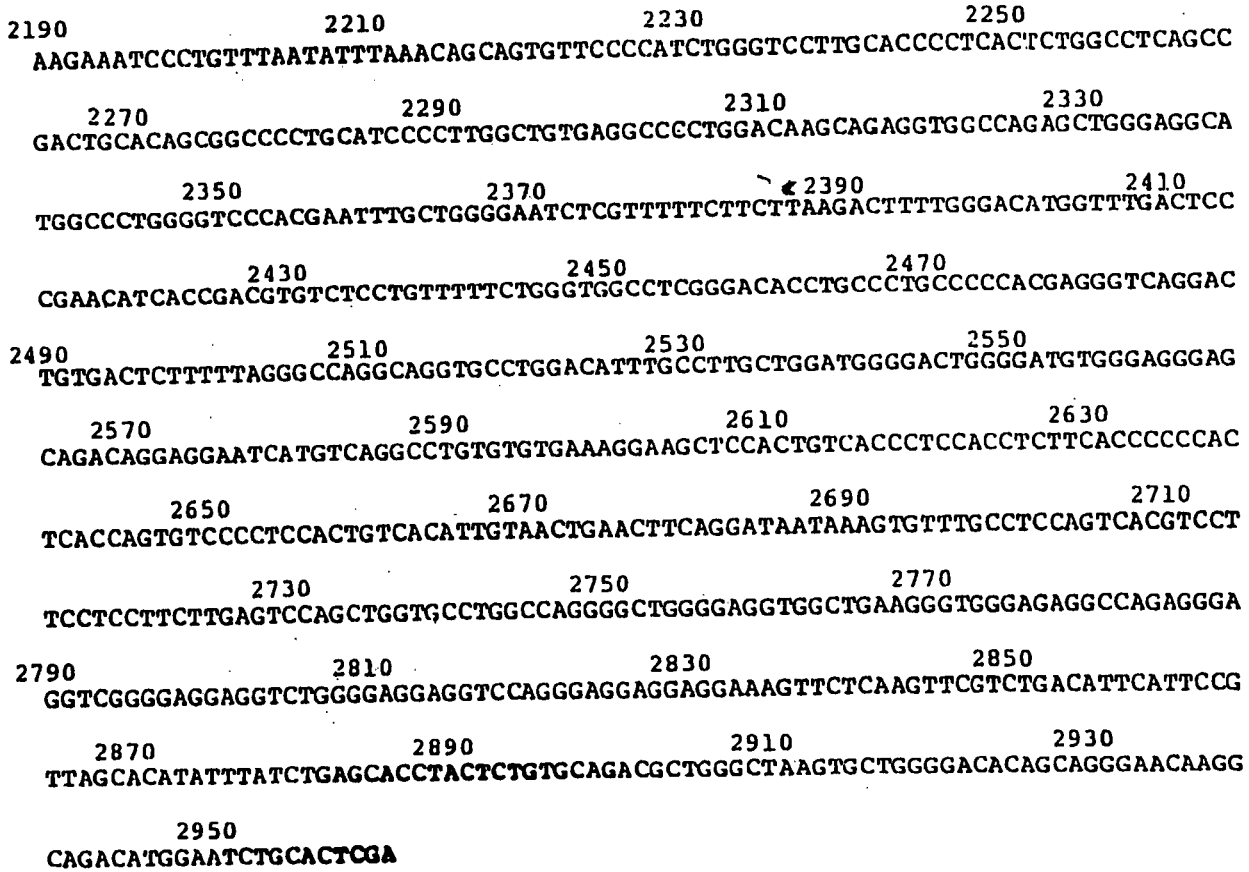

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Fig. 6

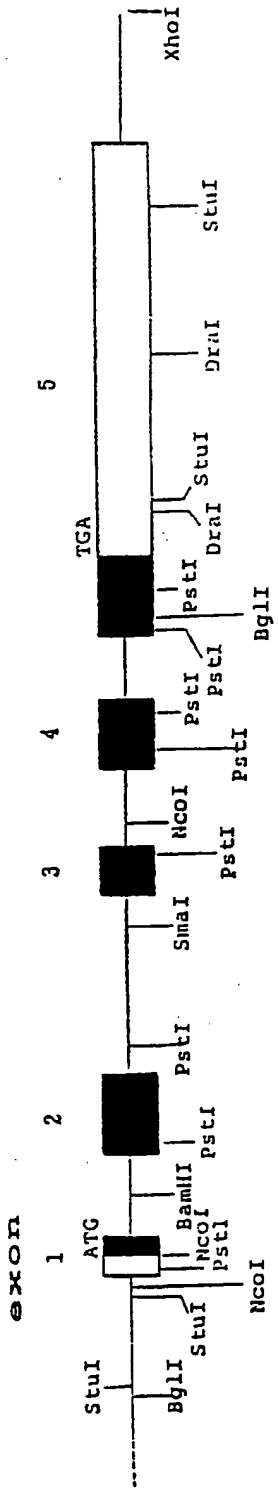


Fig. 7

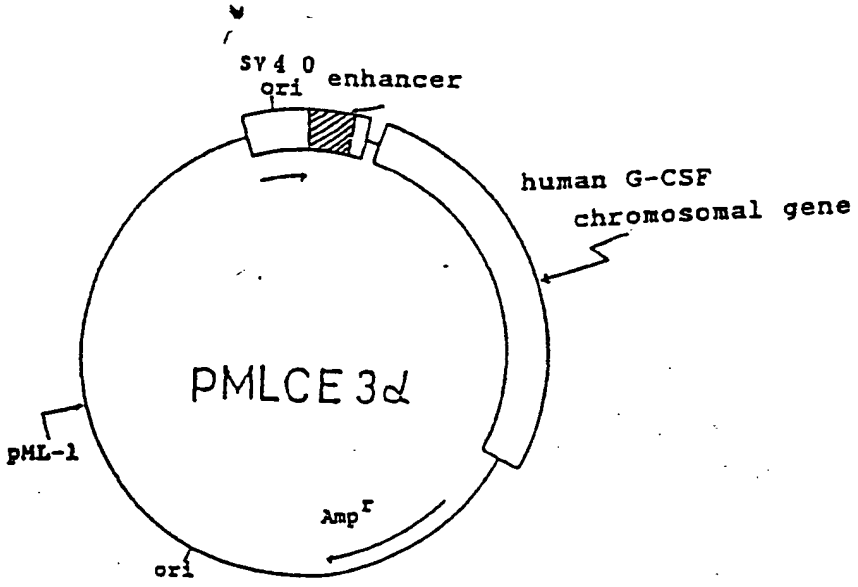

SV40 ori
enhancer
human G-CSF
chromosomal gene
PMLCE 3α
pML-1
Amp^r
ori

Fig. 8

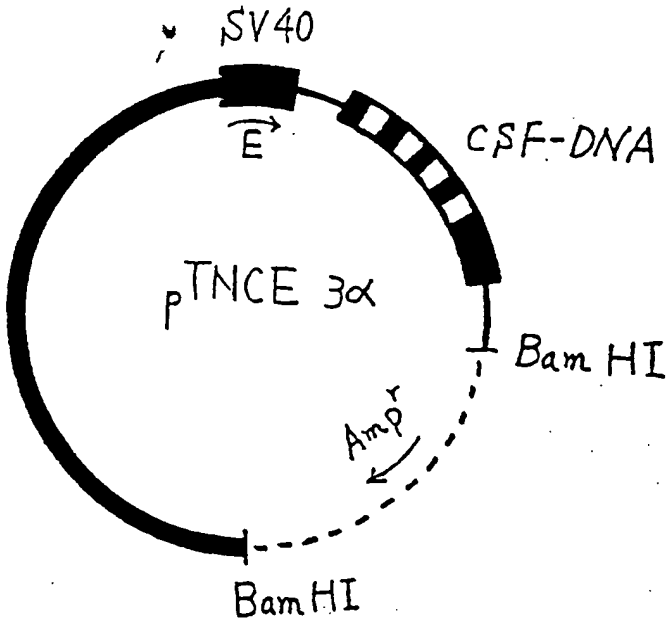
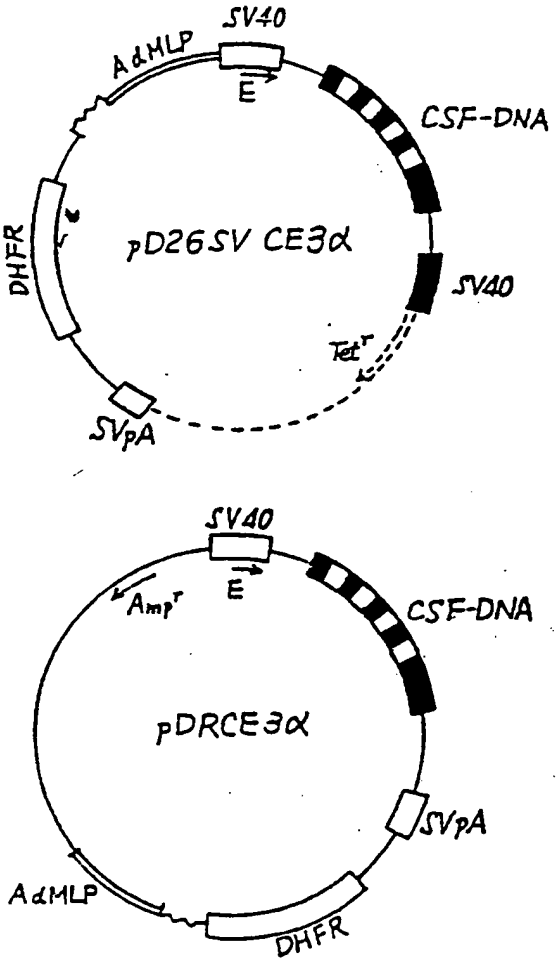


Fig. 9


SV40
AdMLP
E
CSF-DNA
pD26SV CE3α
DHFR
SV40
Tet^r
SVpA
SV40
Amp^r
E
CSF-DNA
pDRCE3α
SVpA
AdMLP
DHFR